



**Radiobiology of Auger electron emitters
and their potential for radionuclide therapy**

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"THE ACTUAL PROBLEMS OF MICROWORLD PHYSICS"
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1. Auger electron emitters – a special class of radionuclides

(physics of Auger effect and physical properties of Auger emitters)

2. Molecular aspects of Auger decay radiobiology

(DNA damage)

3. Auger emitters potential for radionuclide therapy

(dual targeting strategy)



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- each decay event is **unique** (in contrast to α -, β - and γ -emitters)
- **nuclear** and **atomic** processes are involved

Auger effect – named after Pierre Auger

Physics of Auger effect



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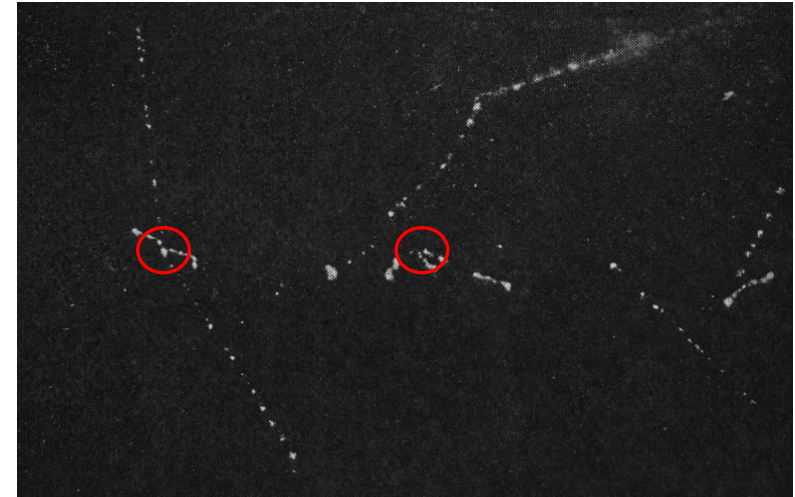
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- Multiple tracks of ionising particles originating from the same point – Auger effect
- Pierre Auger interpretation – simultaneous emission of several electrons from ionised atom

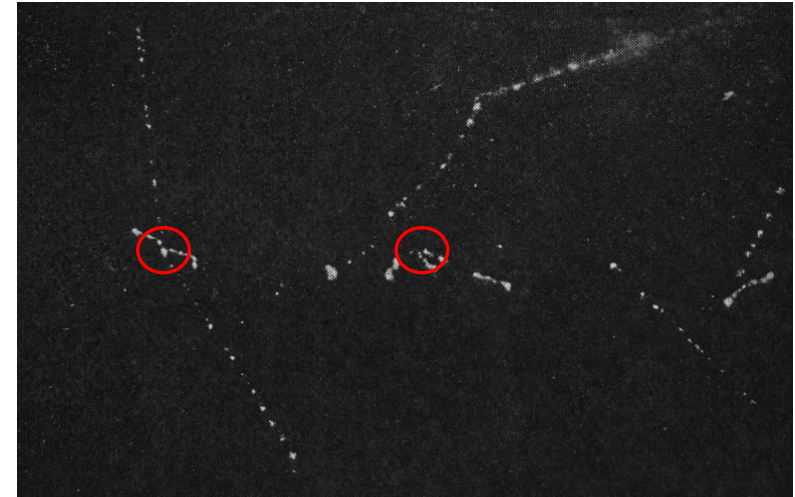
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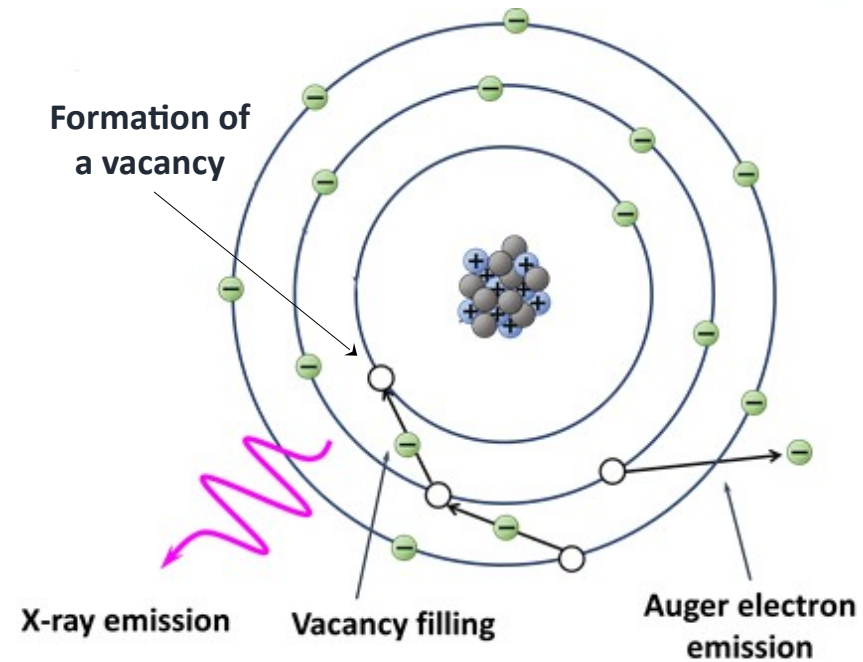


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Main feature of Auger effect – emission of **multiple** electrons (Auger electrons) initiated by a **single** atomic event

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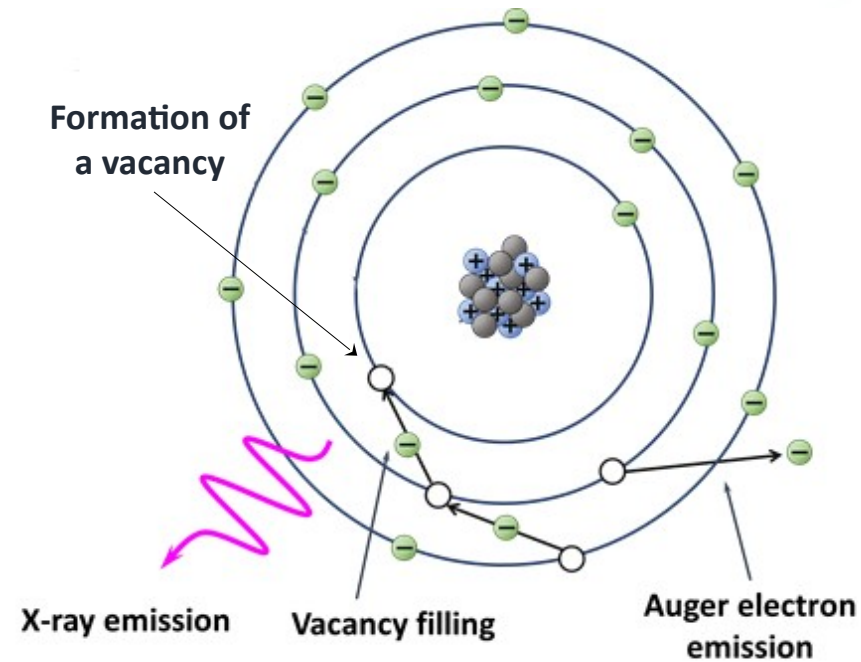
Physics of Auger effect



Auger effect involves:

- **Formation of a vacancy in an inner atomic shell**
- **Filling of this vacancy by another electron; accompanied by**
 - formation of a new vacancy, with the transition energy released as:
 - X-ray photon or
 - emission of another electron with formation of one more vacancy
- **The process of vacancy filling and electron emission (Auger cascade) continues until**
 - all vacancies reach the outer shell (in vacuum) and
 - are filled with electrons from neighbouring atoms (in medium)

Physics of Auger effect



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In summary, Auger effect is:

- emission from an atom of **multiple electrons (Auger electrons)** that is
- initiated by an **inner shell vacancy**

Physics of Auger effect



Inner shell vacancy formation mechanisms:

- photo effect (in Pierre Auger experiments)
- electron capture
- internal conversion

Physics of Auger effect

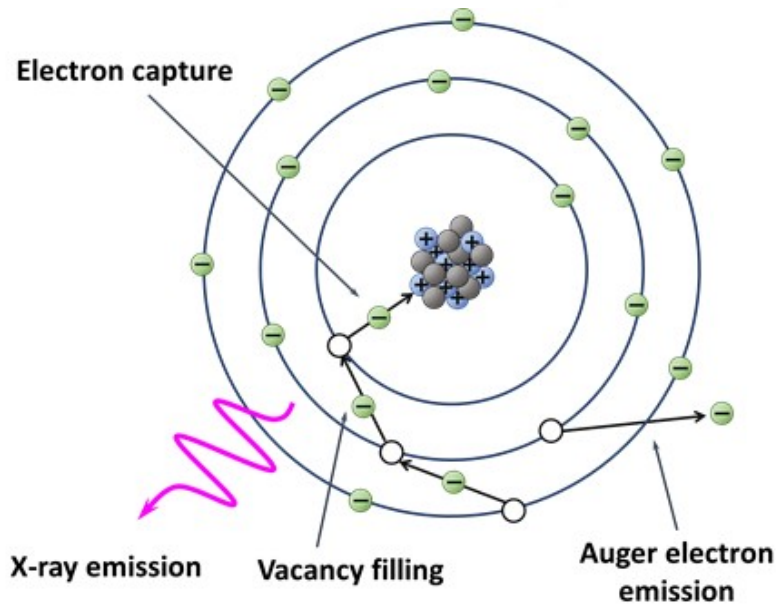


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Electron capture (e-capture, K-capture) – a type of radioactive decay (β -decay)

- capture by a nucleus of an electron from inner shell
- reaction $p^+ + e^- \rightarrow n + \nu$ (a type of β -decay)
- the charge of the nucleus decreases by 1, $Z \rightarrow Z - 1$



Physics of Auger effect



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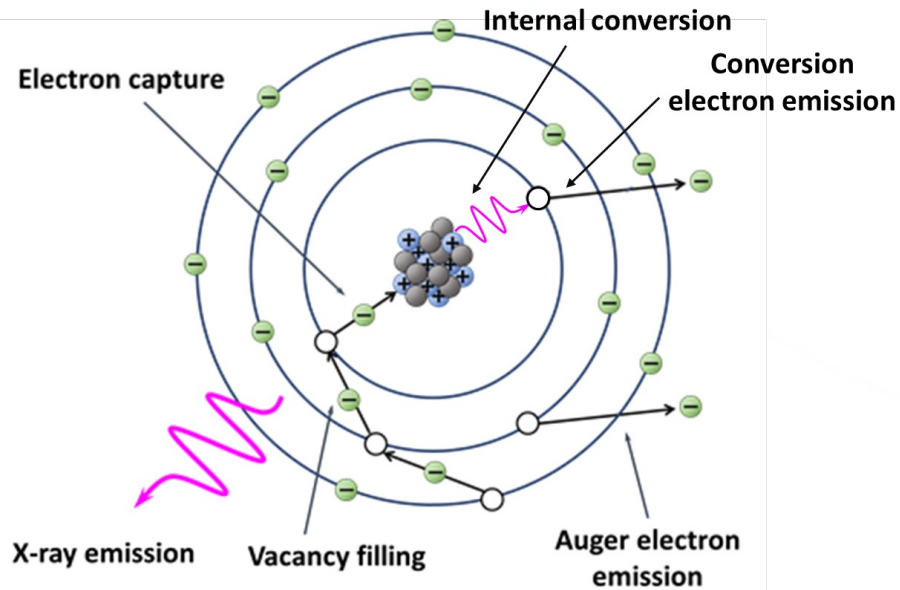
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Internal conversion – a type of radioactive decay

- relaxation of an excited nucleus by transferring energy to the inner shell electron with subsequent emission of this electron



Physics of Auger effect



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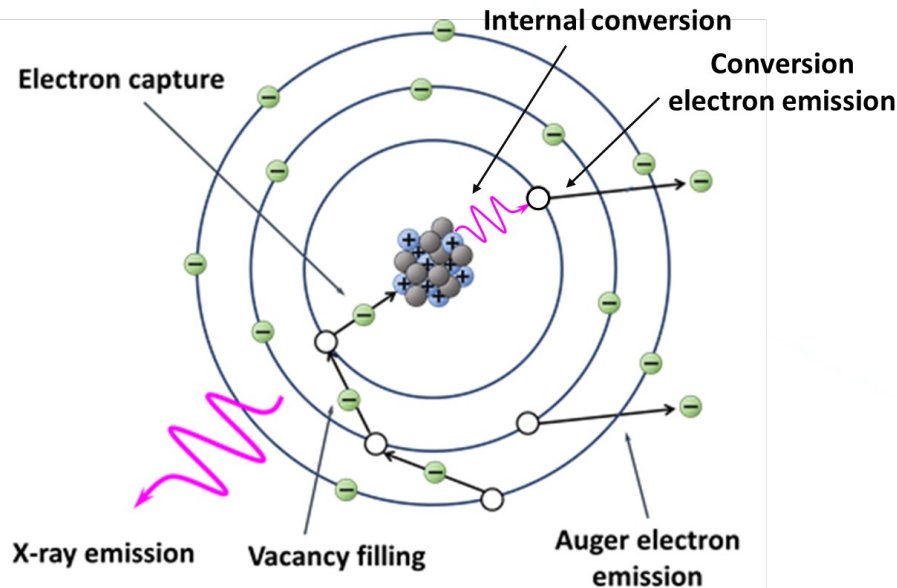
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Internal conversion – a type of radioactive decay

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- Photo effect and formation of a vacancy can occur in any atom (non-radioactive) subjected to photon (X-ray) irradiation
- **Electron Capture and Internal Conversion** – types of radioactive decay –
> properties of a range of radionuclides
- **These radionuclides – Auger electron emitters or Auger emitters**

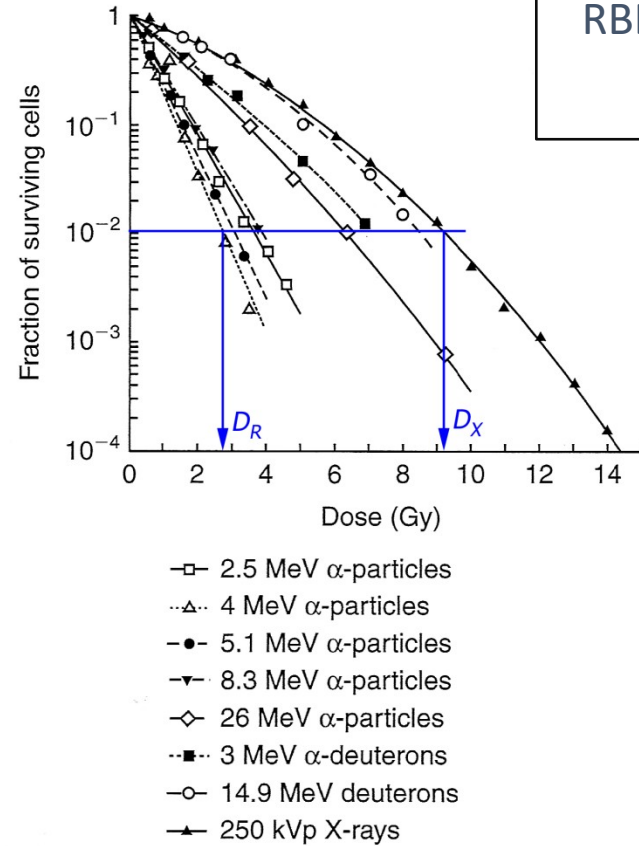
Radiobiological Properties of Radionuclides for Nuclear Medicine



Properties of radionuclides and their relevance for radionuclide endoradiotherapy

Property/Type	β -emitters	α -emitters	Auger emitters
Energy of ionizing particles	0.1 – 1 MeV	A few MeV	?
LET (Linear Energy Transfer)	Low (≈ 1 keV/ μ m)	High (80 -100 keV/ μ m)	?
Range of ionizing particles/Damage localisation	Order of mm/tissue size	Order of 10 μ m /cell size	?
RBE (Relative Biological Effectiveness)	Low (~ 1)	High (~ 3)	High
Availability/Production	Relatively simple	Complex	Relatively simple
Conjugation chemistry	Covalent binding/chelating compounds		

Dose – effect curves and biological effectiveness of radiation



Data from Barendsen, 1968

RBE – the ratio of doses of the standard and tested radiation that result in the same level of biological effect

$$RBE = \frac{D_x}{D_R}$$

D_x – dose of the standard radiation (x-rays)
 D_R – dose of the tested radiation

For 4 MeV α-particles RBE = 3.2

➤ What quantitative property of ionising radiation determines its RBE and therefore allows to predict biological response?

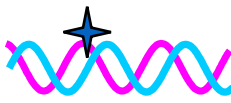
➤ Ionisation density

X-rays – sparsely ionising radiation

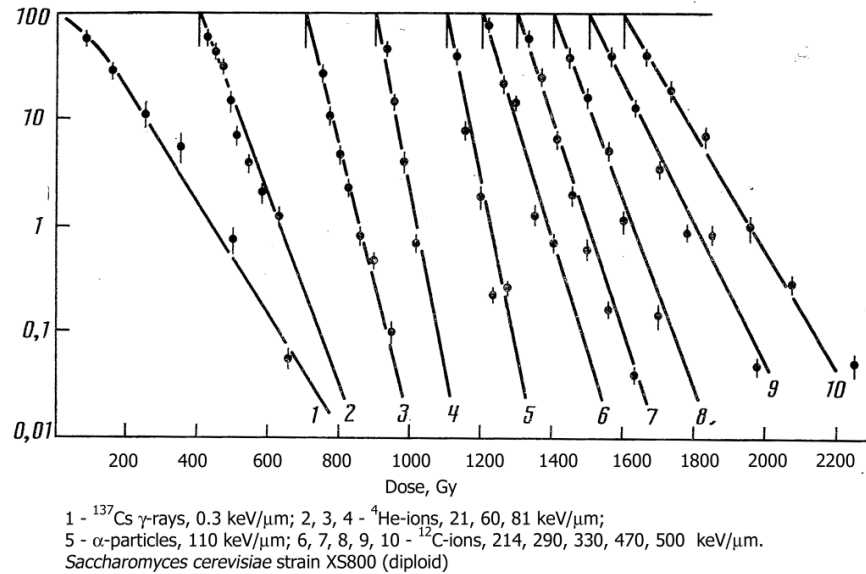
α-particles – densely ionising radiation

Introduction of LET – Linear Energy Transfer
(a measure of ionisation density)

RBE-LET relationship

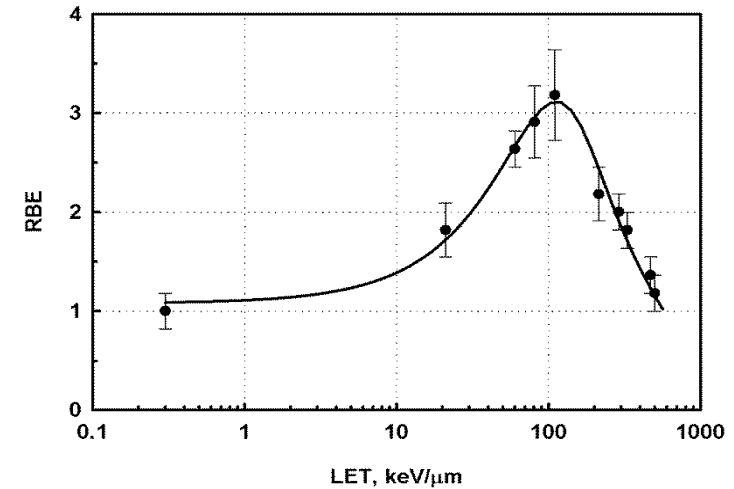


How Relative Biological Effectiveness depends on LET of radiation?



Data from P. Lobachevsky et al, 1988
obtained at U200, LNR JINR

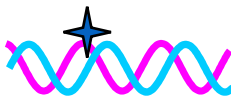
$$RBE = \frac{D_x}{D_R}$$



RBE-LET relationship is complex:

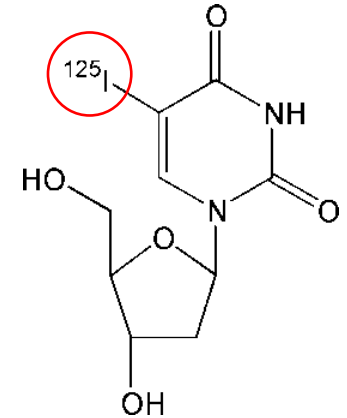
- Increasing RBE in the range up to ~ 100 keV/ μm
- Maximum at ~ 100 keV/ μm
- Fast decrease at LET above ~ 100 keV/ μm

Biological consequences of Auger decay - DNA damage



Early radiobiological experiments with ^{125}I

- model – bacteriophage T1 and T4
- end points – survival and induction of single strand and double strand DNA breaks (SSB and DSB)
- radionuclide carrier - [^{125}I] iodo-deoxyuridine (5- ^{125}I UdR), thymidine analogue, DNA synthesis precursor
- covalent bond of Auger emitter with DNA

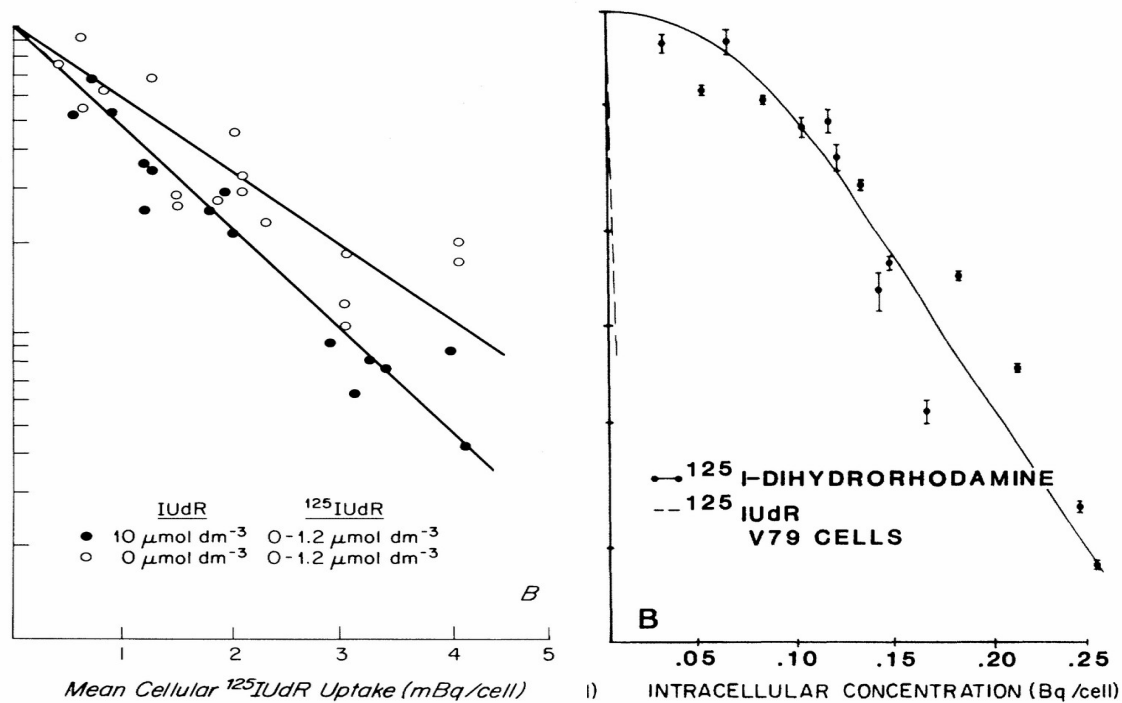
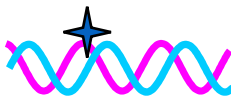


Major conclusion

- decay of ^{125}I , covalently incorporated into DNA results in formation of DNA **DSB with a probability close to 1**
- one DNA DSB induced by ^{125}I -decay is a lethal event for bacteriophage

Schmidt & Hotz, IJRB, 1973
Krisch & Ley, IJRB, 1974
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Radiation biology of Auger decay – cellular effects



Localisation of Auger emitter relative to DNA is of critical importance

Experiments with mammalian cells (*Kassis & Adelstein, Rad Res, 1987*)

Effect of ^{125}I decay: extracellular, in cytoplasm, in cell nucleus

Decay of DNA incorporated ^{125}I ~ 6 fold more efficient than decay of ^{125}I in cytoplasm (based on absorbed dose)

Carrier of ^{125}I	Localisation	D_{37} mBq/cell	D_{37} Gy
Na^{125}I	Extracellular	-	-
$[^{125}\text{I}]$ -iodo-dihydrorhodamine (^{125}I -DR)	Cytoplasm	109	4.62
$[^{125}\text{I}]$ -iodo-deoxyuridine ($5\text{-}^{125}\text{I}$ UdR)	Nucleus (DNA)	1.30	0.80

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LET and RBE of Auger electrons – is LET of Auger electrons high?



- It is quite common to attribute high radiobiological efficiency (RBE) of Auger decay to high LET of Auger electrons
- LET of Auger electrons cited as 4 – 26 keV/μm, which is considered as high LET

Citation: “Both α-particle and Auger electron emitters are considered as high LET radiation qualities; with an LET of 80–100 keV/μm for α-particle emitters and an LET of 4–26 keV/μm for Auger electron emitters.”

Fourie H et al. Estimating the Relative Biological Effectiveness of Auger Electron 123 Emitter I in Human Lymphocytes. Front Phys. 2020;8:567732.

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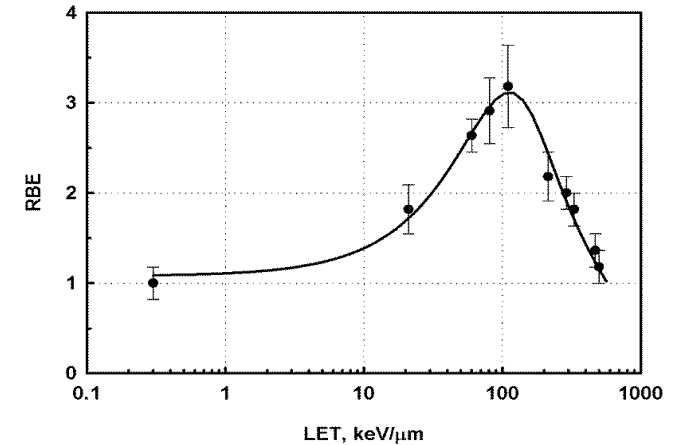
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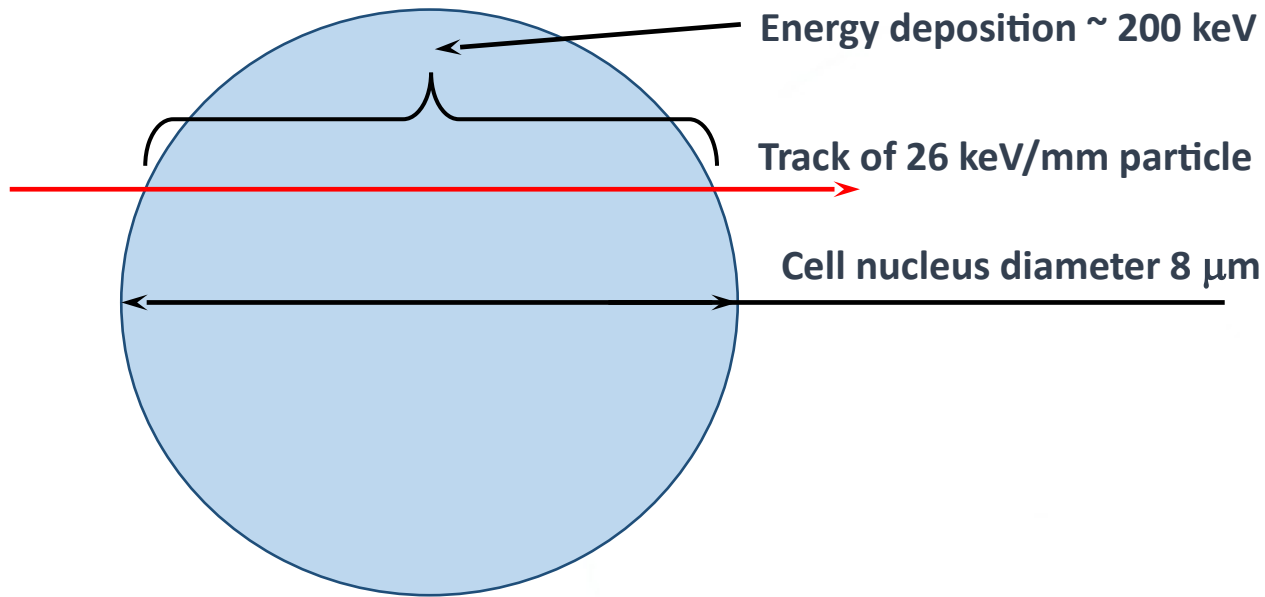
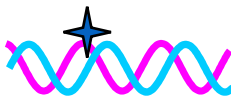
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Electron energy, eV	Range, μm(nm)	dE/dx, keV/μm
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60	0.0038 (3.8)	26
100	0.0054 (5.4)	26
200	0.0094 (9.4)	23
400	0.019 (19)	17
600	0.032 (32)	14,5

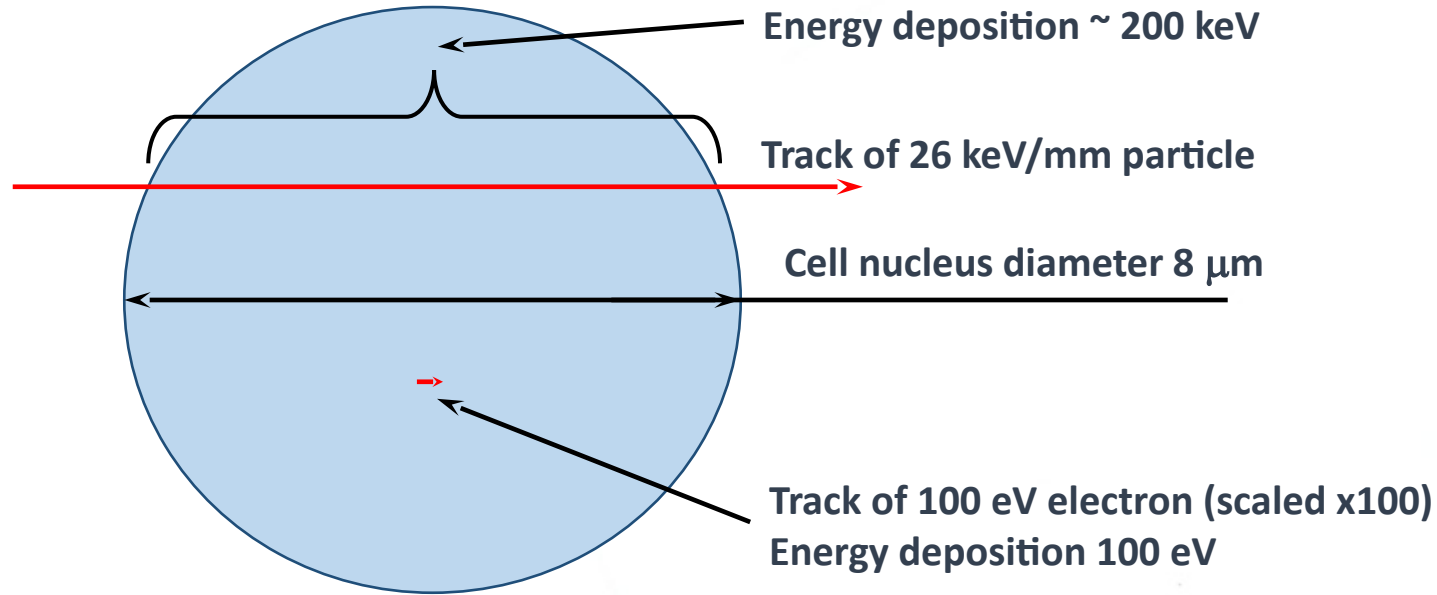
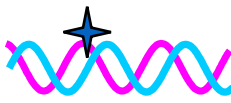
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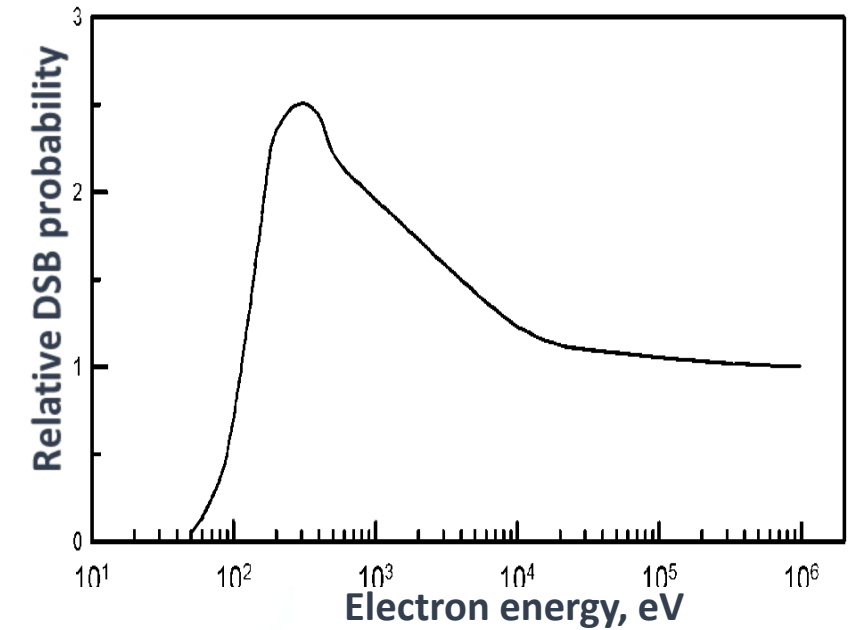
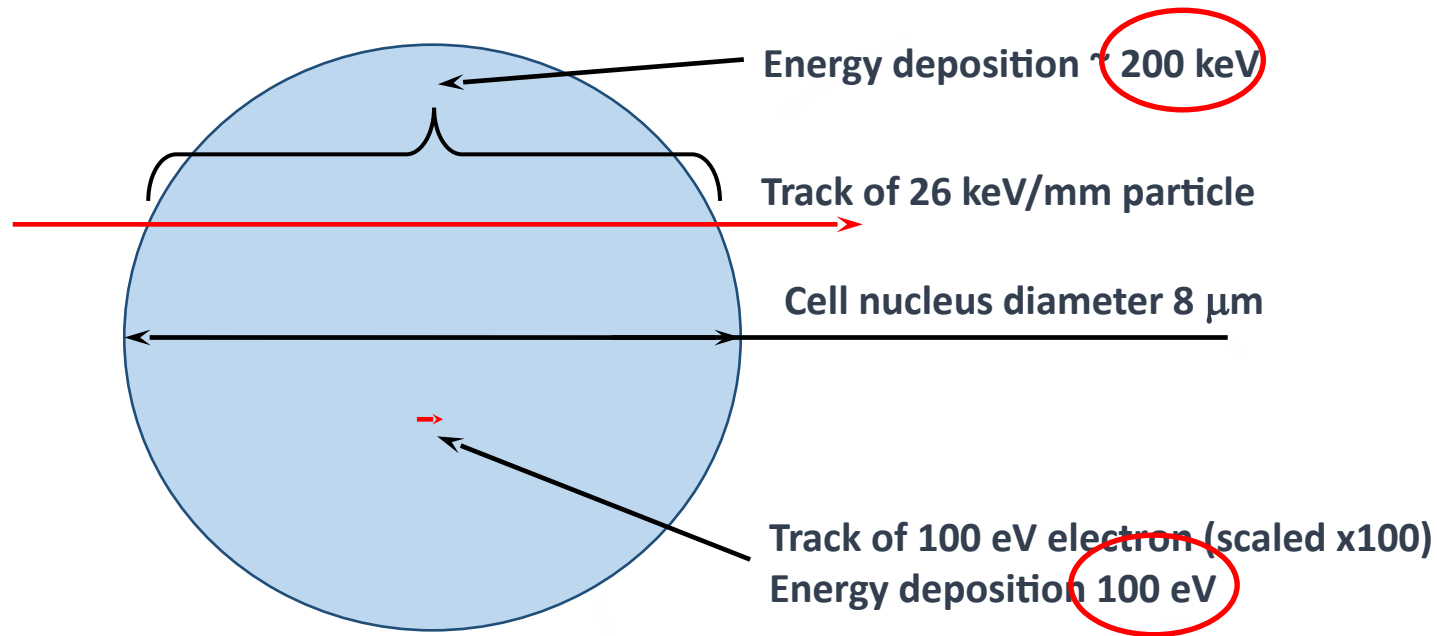
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LET and RBE of Auger electrons – is LET of Auger electrons high?



Michalik & Frankenberg, 1994

- Are these two events biologically equivalent? - No
- The concept of **LET is not applicable** to Auger electrons (in radiobiological context)
- LET is a **macroscopic** (statistical average) value
- **Microscopic** energy deposition analysis is required
- High **local energy** deposition – result of simultaneous emission of **multiple** electrons



Important physical properties of Auger emitters for radiation biology:

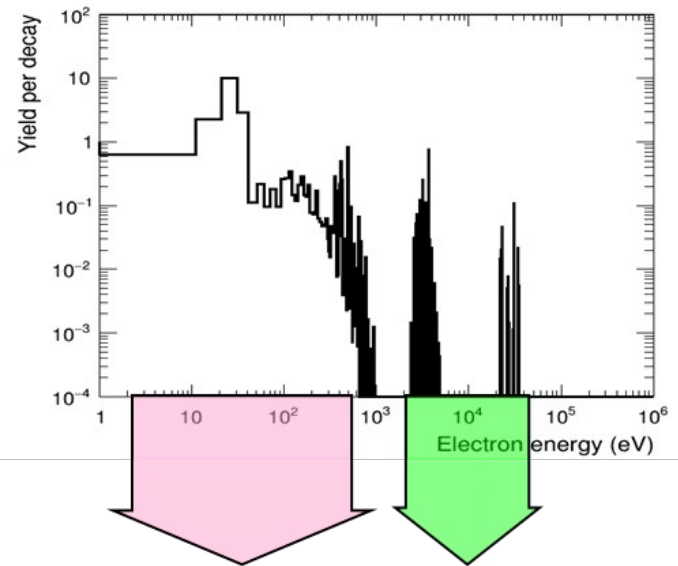
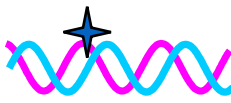
- the **energy** and **range** of Auger electrons
- the **number** of electrons emitted per decay



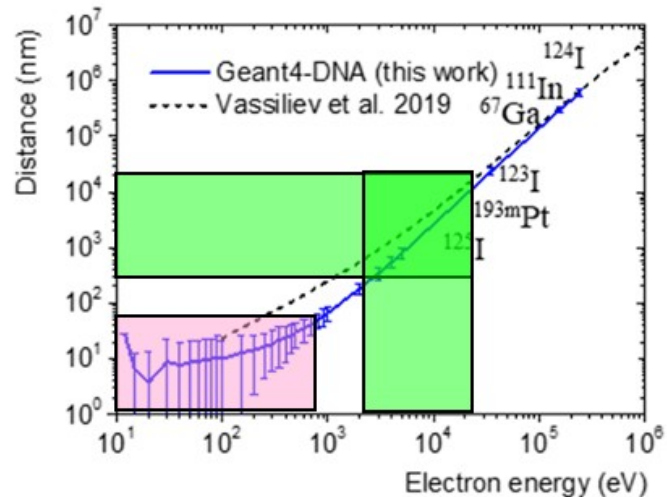
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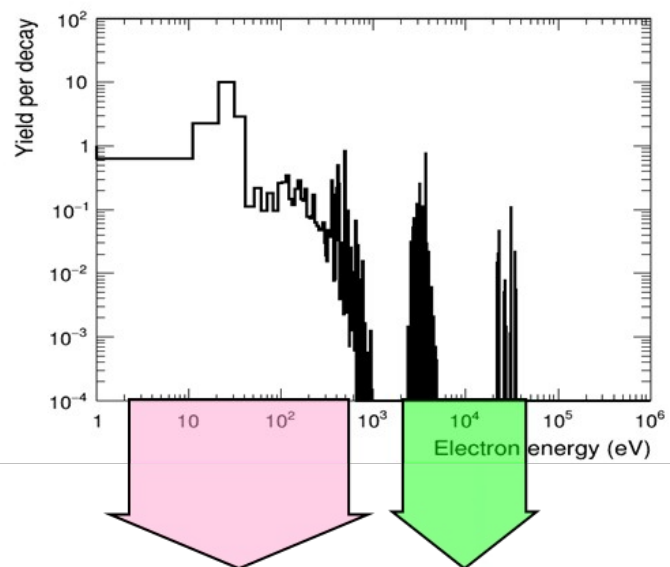
Auger emitters – energy and range of Auger electrons (^{125}I)



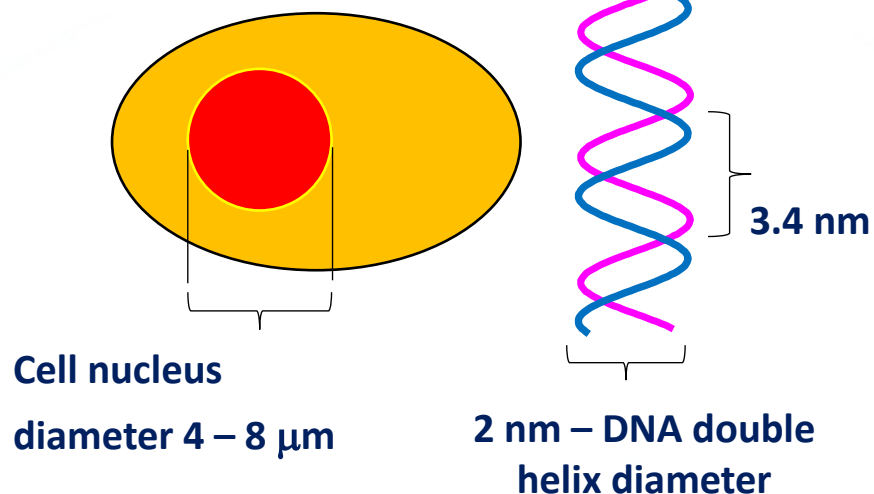
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1 – 50 nm **0.2 - 20 μm**



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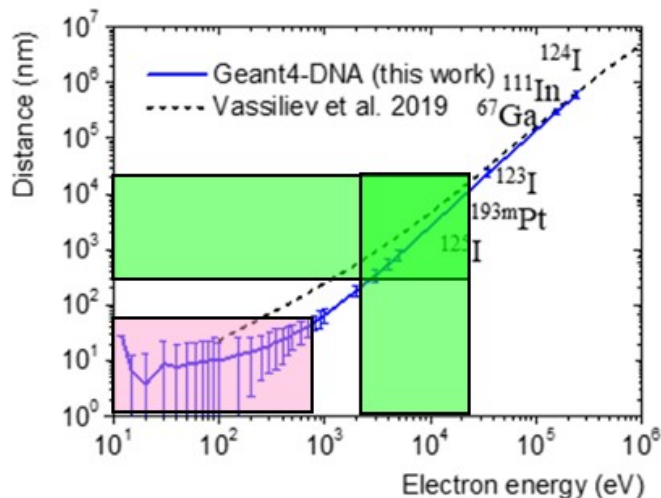


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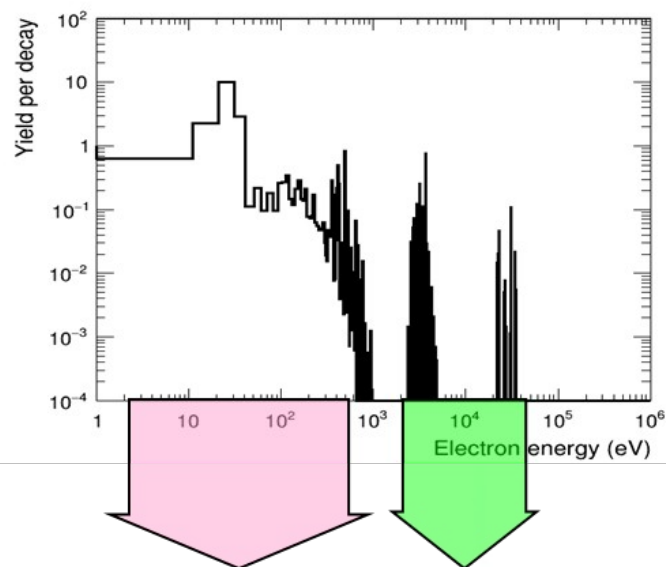


Range of Auger electrons:

- for energies < 1 keV – up to 50 nm, dimensions of DNA molecule (2 nm – DNA double helix diameter, 0.3 nm – inter-base distance)
- for energies < 30 keV – up to 20 μm , nucleus size (8 μm – nucleus diameter in mammalian cells)

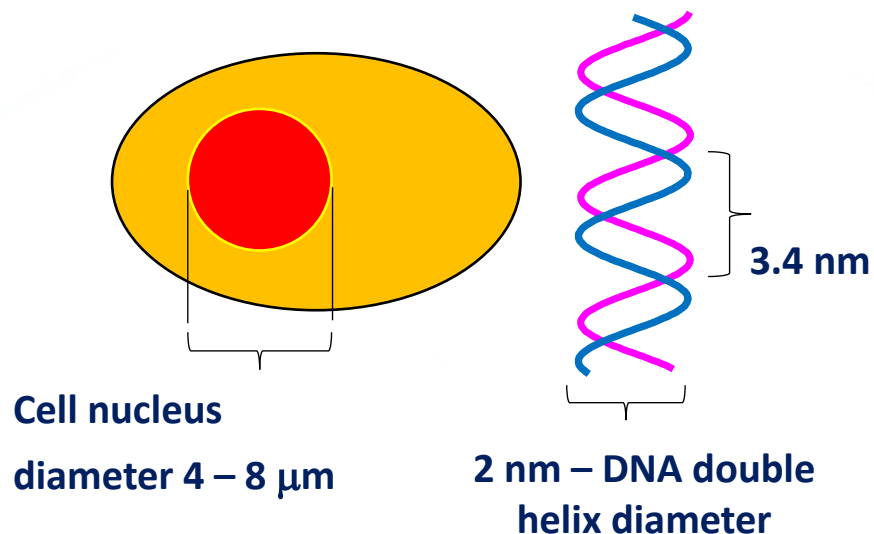
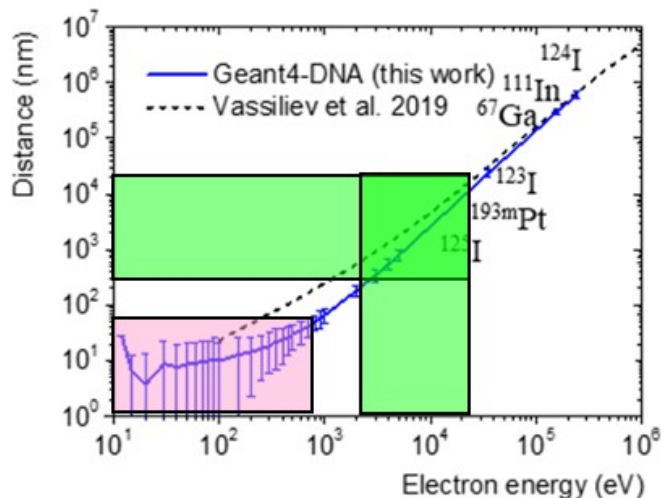


Auger emitters – energy and range of Auger electrons (^{125}I)



0.02 - 0.9 keV
1 - 50 nm

2.5 - 35 keV
0.2 - 20 μm



Number of Auger electrons by ^{125}I decay –
25 on average

Range of Auger electrons:

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- for energies < 30 keV – up to 20 μm , nucleus size (8 μm – nucleus diameter in mammalian cells)

Critical consequences of Auger decay for radiation biology:

- the range of Auger electrons is comparable to the size of cell nucleus
- high local deposition of absorbed energy from multiple electrons (within the size of DNA and cell nucleus)



Important physical properties of Auger emitters for radiation biology:

- the energy and range of Auger electrons
- the **number** of electrons emitted per decay

Auger emitters – biologically relevant properties



Nuclide	Total Decay Energy keV	Half-Life Time	Auger electrons	Conversion electron	X-Rays	γ -Rays
			Yield/Decay (Energy, keV)	Yield/Decay (Energy, keV)	Yield/Decay (Energy, keV)	Yield/Decay (Energy, keV)
⁶⁷ Ga	201.6	3.26 d	4.7 (6.26)	0.32 (28.1)	0.57 (4.9)	0.88 (162.3)
¹¹¹ In	419.2	2.80 d	14.7 (6.75)	0.16 (26.0)	0.89 (20.0)	1.84 (366.5)
^{99m} Tc	142.6	6.01 h	4.0 (0.90)	1.1 (15.4)	0.079 (1.37)	0.89 (125.0)
¹²³ I	200.4	13.2 h	14.9 (7.4)	0.15 (20.2)	0.93 (24.1)	0.87 (148.6)
¹²⁴ I*		4.18 d	8.4 (4.74)	0.0035 ()	0.65 (17.1)	0.92 (863)
¹²⁵ I	61.4	60.4 d	24.9 (12.2)	0.94 (7.2)	1.53 (39.7)	0.065 (2.3)

Howell RW, *Medical Physics*, 1992

Pomplun E, 1992

Humm JL, 1994

Lee BQ, IJRB, 2015

* Positron emission yield 0.23



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(physics of Auger effect and physical properties of Auger emitters)

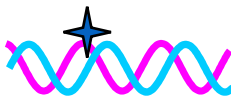
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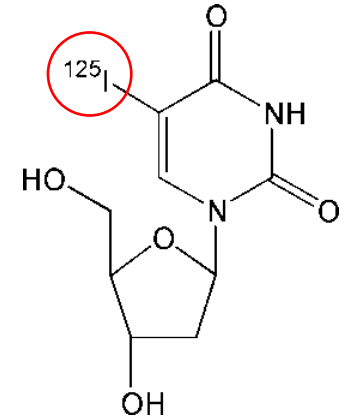
(dual targeting strategy)

Biological consequences of Auger decay - DNA damage



Early radiobiological experiments with ^{125}I

- model – bacteriophage T1 and T4
- end points – survival and induction of single strand and double strand DNA breaks (SSB and DSB)
- radionuclide carrier - [^{125}I] iodo-deoxyuridine (5- ^{125}I UdR), thymidine analogue, DNA synthesis precursor
- covalent bond of Auger emitter with DNA



Major conclusion

- decay of ^{125}I , covalently incorporated into DNA results in formation of DNA **DSB with a probability close to 1**
- one DNA DSB induced by ^{125}I -decay is a lethal event for bacteriophage

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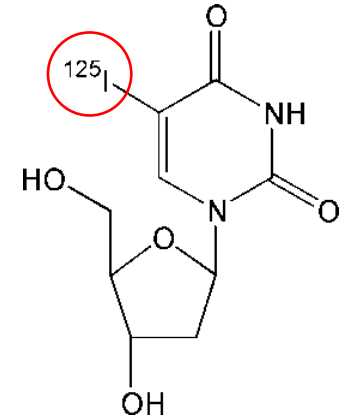


^{125}I : One decay event – one DNA DSB – 100% efficiency of DNA damage

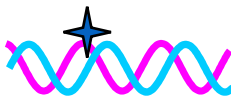
Mechanisms?

- 1. Auger electrons emission (obvious and traditional)**
- 2. Non-traditional mechanism: (not obvious and alternative)**

Molecule fragmentation as a result of coulombic explosion of multiply charged (+7-20) daughter atom ^{125}Te



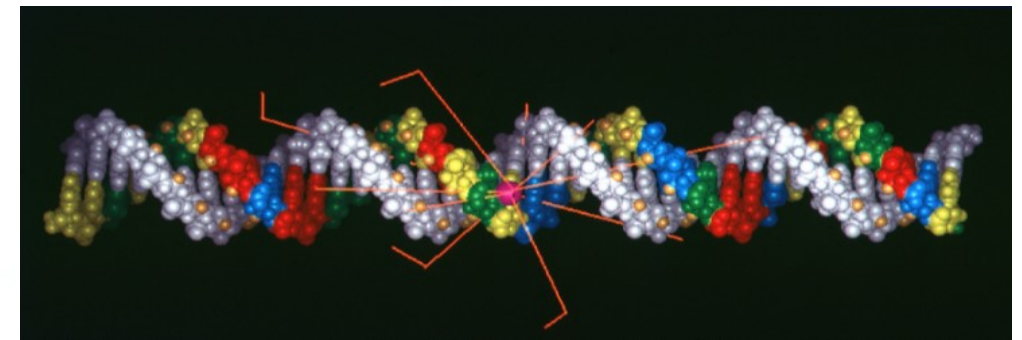
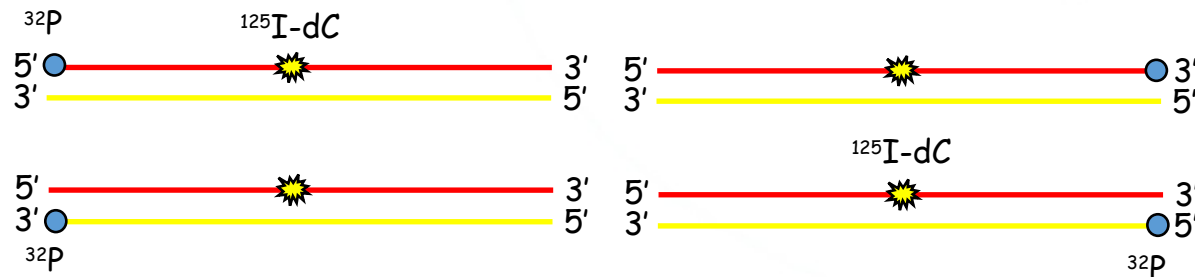
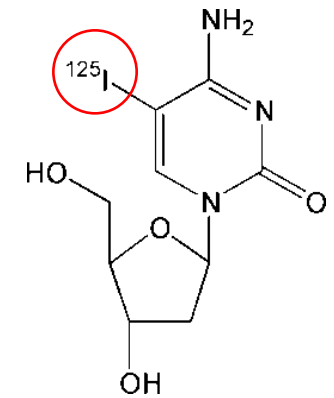
Auger decay DNA damage – mapping DNA breaks



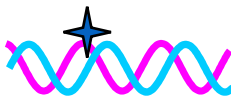
Experiments with synthetic oligodeoxynucleotides

(Lobachevsky & Martin, Rad Res, 2000)

- model – 41 bp oligodeoxynucleotide incorporating single ^{125}I and complementary 41 bp fragment
- end point – induction of DNA SSB +/- DMSO - dimethylsulphoxide
- method – sequencing gel electrophoresis (^{32}P -label for detection)
- radionuclide carrier – [^{125}I]-iodo-deoxycytidine (5- ^{125}I dC), incorporated in one DNA strand

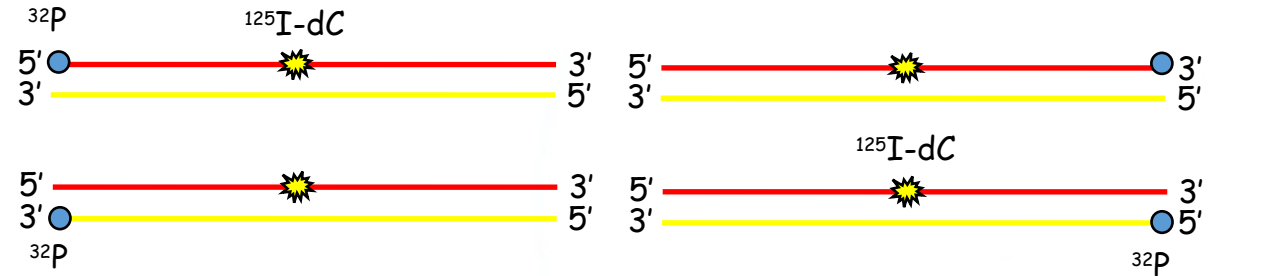


Auger decay DNA damage – mapping DNA breaks

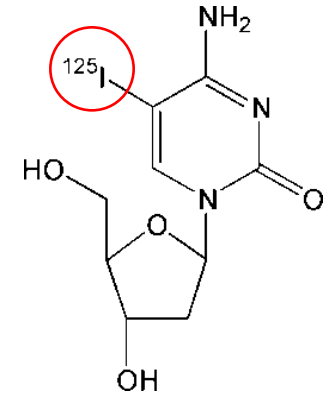
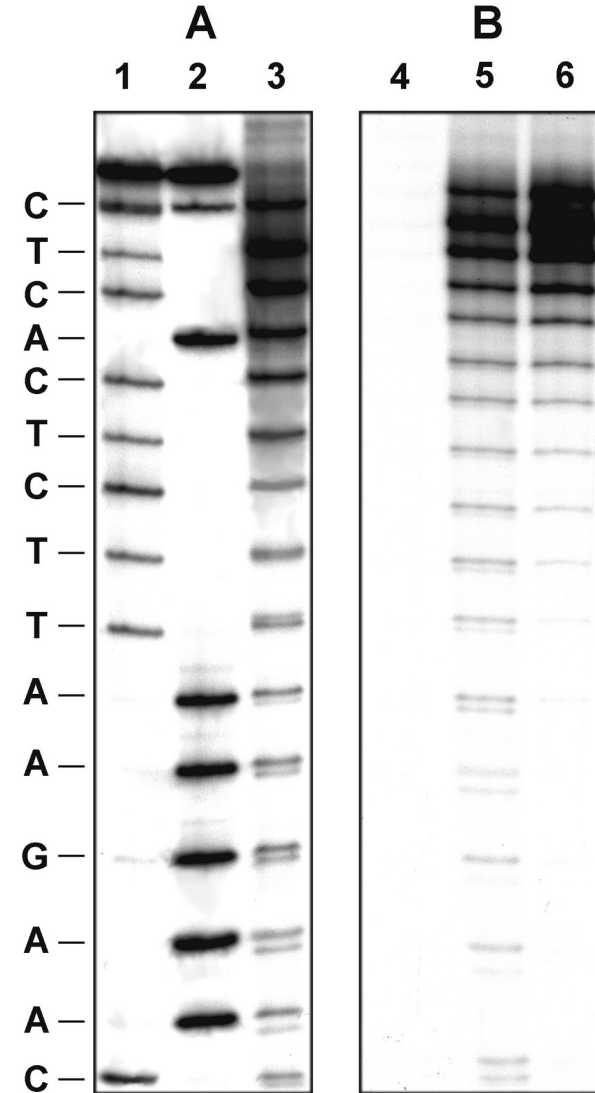


Experiments with synthetic oligodeoxynucleotides

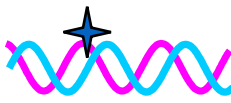
Detection of DNA fragments



Decay induced
fragments of various
length

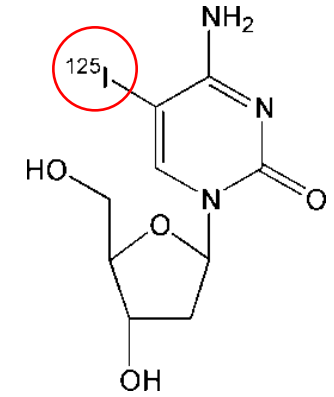
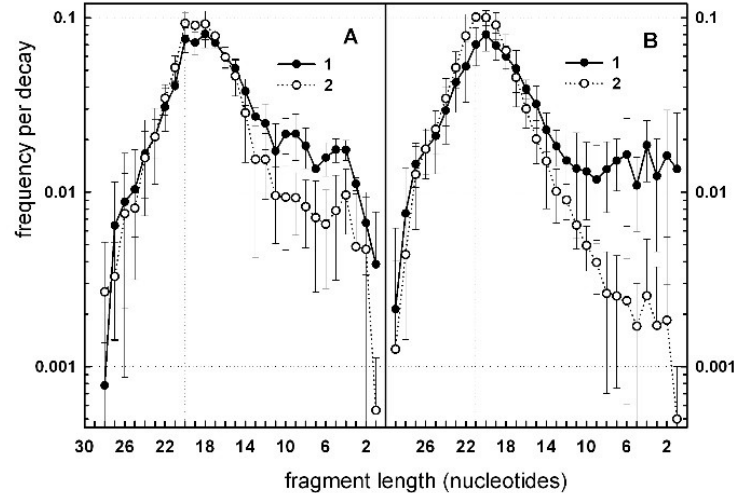
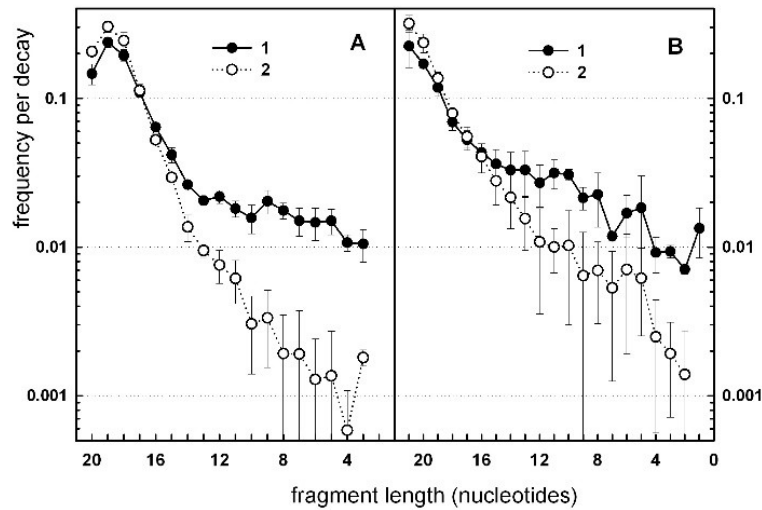


Auger decay DNA damage – mapping DNA breaks



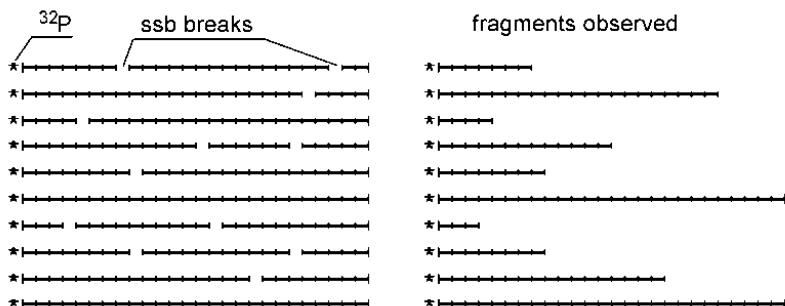
Experiments with synthetic oligodeoxynucleotides

Fragment size distributions



$$f_i = p_i(1 - p_{i-1}) \dots (1 - p_0)$$

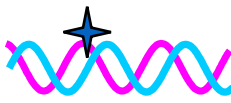
$$p_i = \frac{1}{1 - \sum_j f_j}$$



f_i – fragment detection frequency

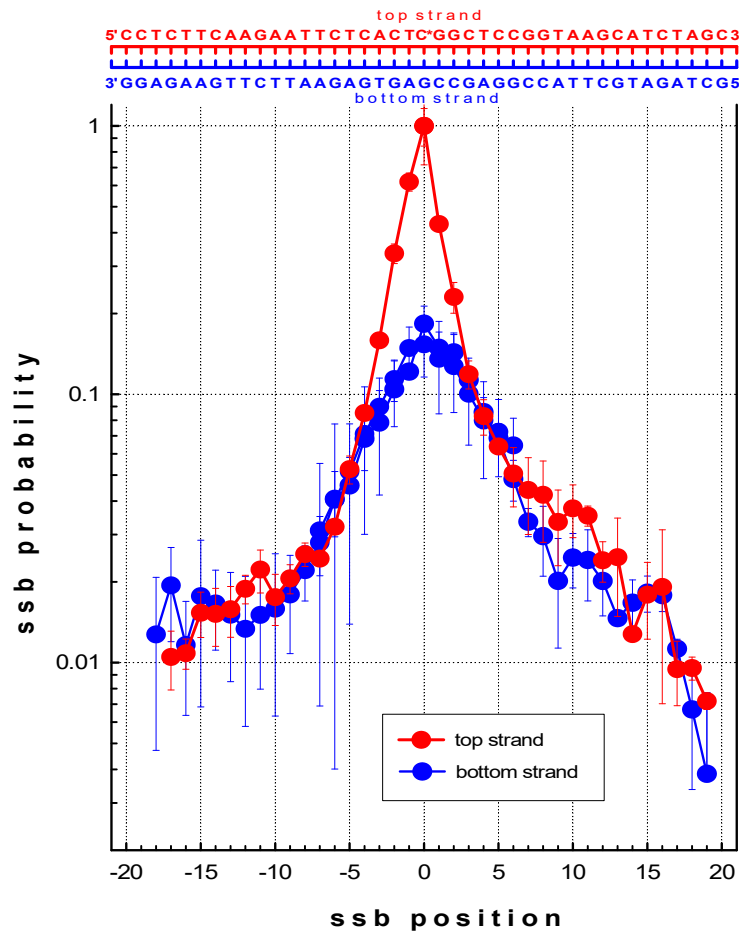
p_i – probability of SSB

Auger decay DNA damage – mapping DNA breaks

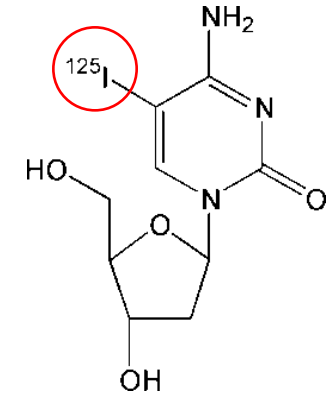
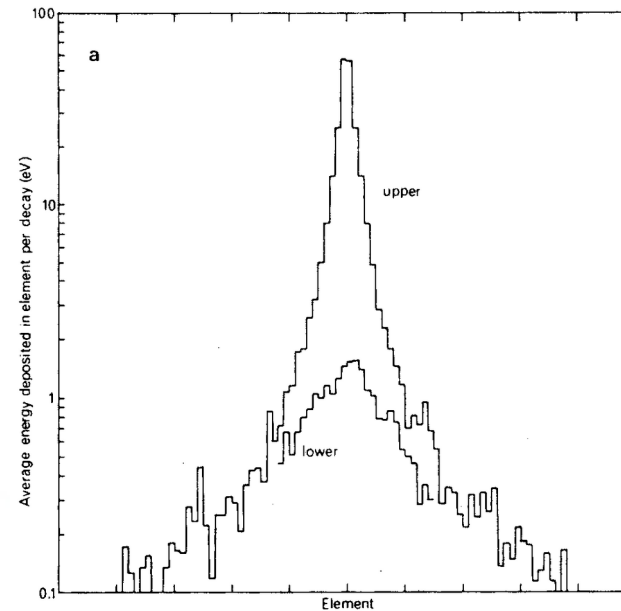
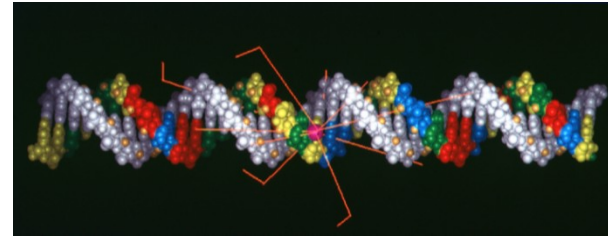


Experiments with synthetic oligodeoxynucleotides

SSB probabilities



DNA SSB per ^{125}I decay probability map



Energy deposition in DNA elements per ^{125}I decay

Humm & Charlton, 1988

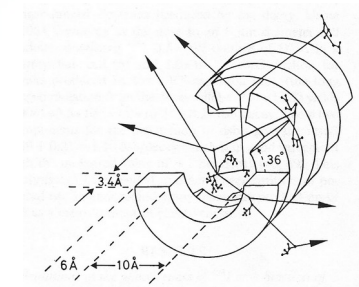
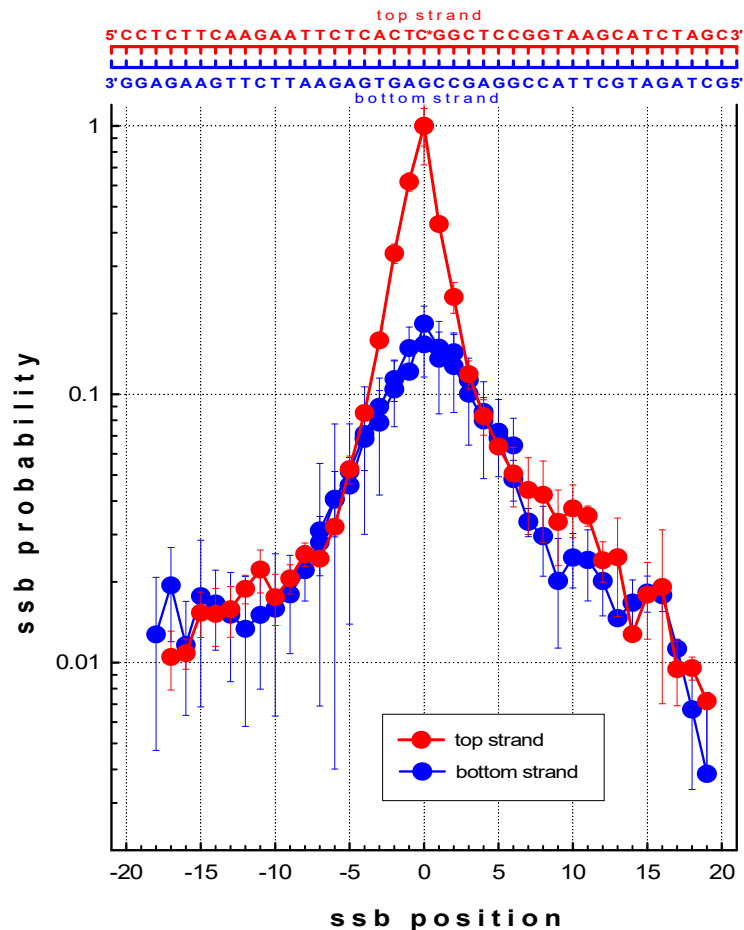
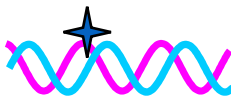


Fig. 1. Schematic diagram of an Auger source decaying in the DNA. The DNA model is described in the text. Each decay produces a set of individual electron tracks producing energy depositions at random through the DNA.

Auger decay DNA damage – mapping DNA breaks

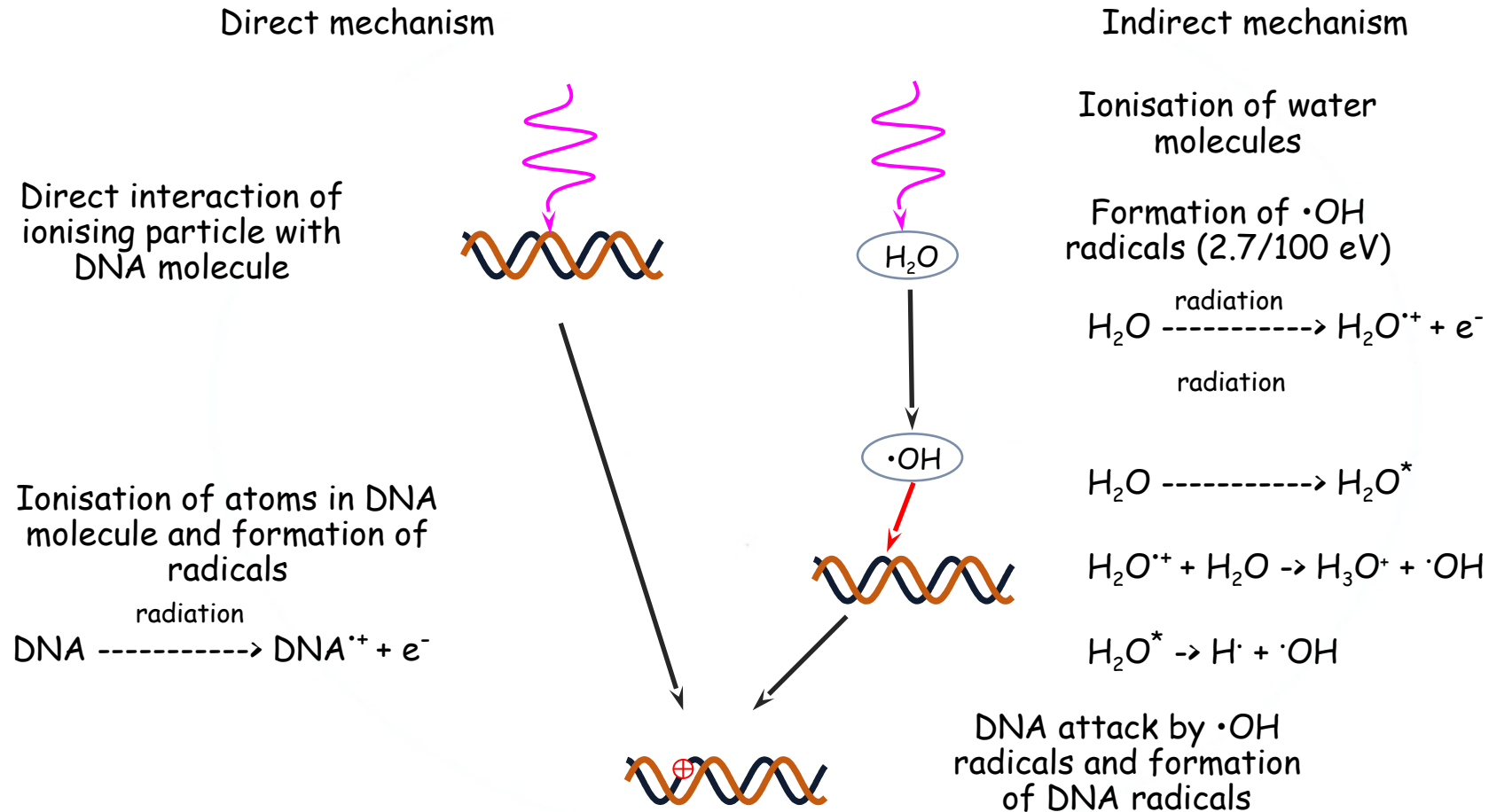
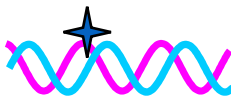


Experiments with synthetic oligodeoxynucleotides

- Majority of breaks within +/- 5 bp
- Probability of DSB – 0.8
- On average 3.67 breaks in top strand and 1.53 breaks in complementary strand
- **Formation of clustered damage – DNA DSB of high complexity – high radiobiological efficiency of ^{125}I -decay**

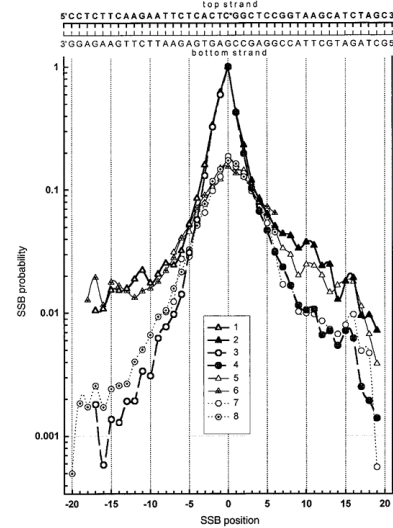
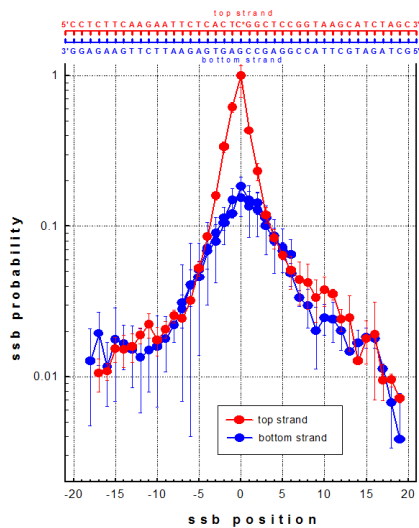
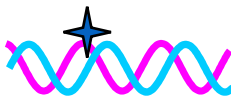
DNA SSB per ^{125}I decay probability map

Direct and indirect mechanism or radiation action



Indirect mechanism can be blocked by addition of radical scavengers, such as DMSO - dimethylsulphoxide

Auger decay – contribution of three DNA damage mechanisms



Contribution of three mechanisms:

- Non-radiation mechanism - neutralisation of charged ^{125}Te atom
- Radiation direct mechanism – direct energy deposition in DNA from Auger electrons
- Radiation indirect – (hydroxyl) radical attack of DNA

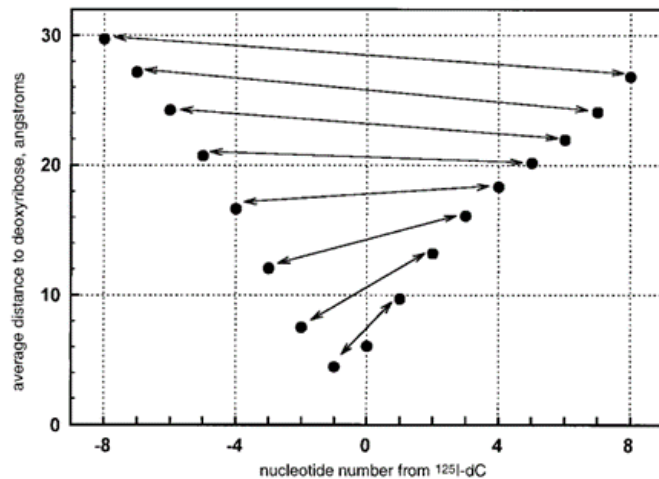
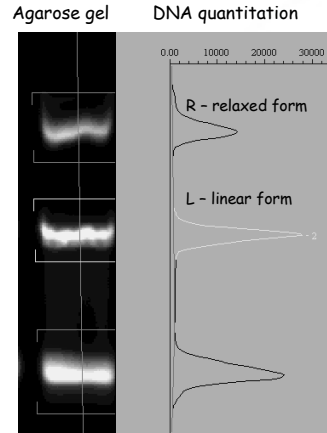
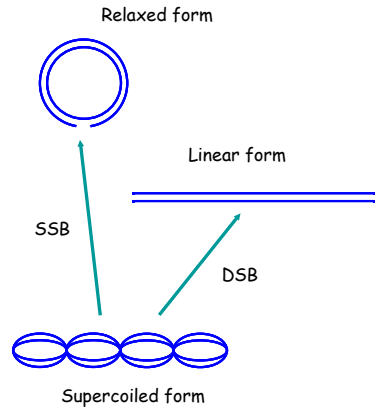
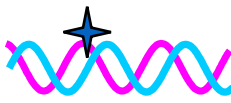


FIG. 1. Average distance from an ^{125}I atom to carbons of the deoxy-ribose moiety in the top strand. Negative nucleotide numbers correspond to the 5' direction, positive ones to the 3' direction. Corresponding nucleotides are indicated by arrows.

Component/DNA strand	Non-radiation (charge neutralisation)	Radiation direct	Radiation indirect (DMSO scavengeable)
^{125}I -incorporating (top)	2.37 (55%)	1.24 (29%)	0.68 (16%)
Complementary (bottom)	0.65 (35%)	0.75 (41%)	0.43 (24%)
DNA duplex	3.02 (49%)	1.99 (33%)	1.11 (18%)

DNA damage – Auger decay in DNA ligand



$$N_{dsb} = \frac{L}{1-L}$$

$$N_{ssb} = \ln\left(\frac{1-L}{S}\right)$$

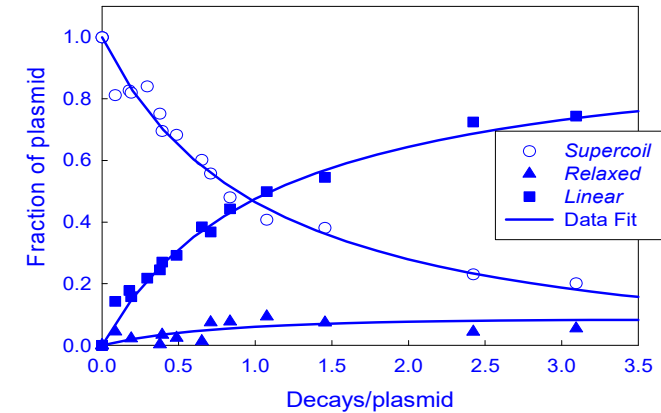
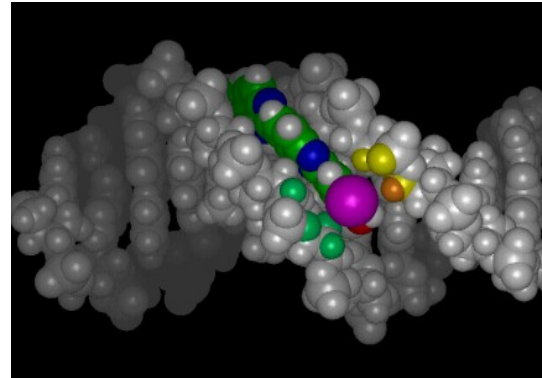
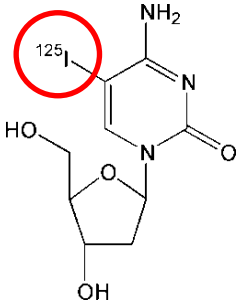
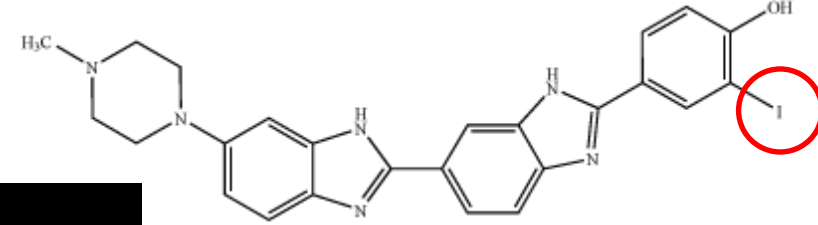
$$S = \frac{e^{-N_{ssb}}}{1+N_{dsb}}$$

$$R = \frac{1-e^{-N_{ssb}}}{1+N_{dsb}}$$

$$L = \frac{N_{dsb}}{1+N_{dsb}}$$

Quantitation of SSB and DSB yield from the fractions of relaxed and linear form in plasmid DNA model
pBR322 – 4361 bp

DNA minor groove binding ligand ¹²⁵I – iodoHoechst 33258

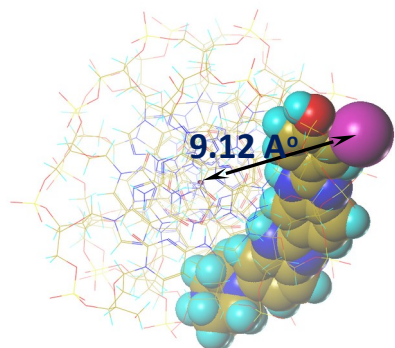


DSB probability – 0.8-0.9 per decay per molecule

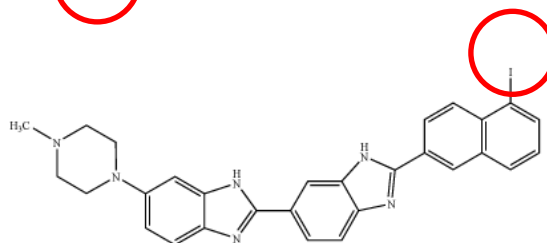
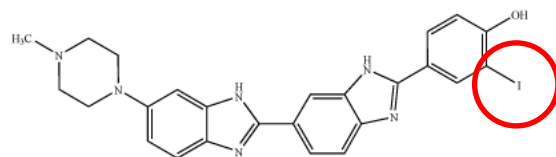
Auger decay in DNA ligand – the role of distance



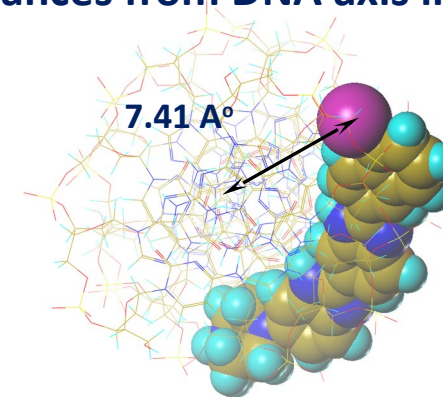
A series of DNA minor groove ligands that position ^{125}I at various distances from DNA axis in plasmid DNA model



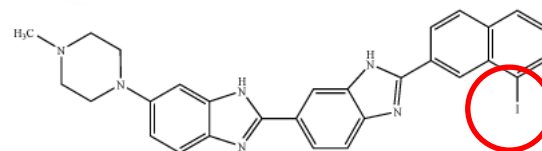
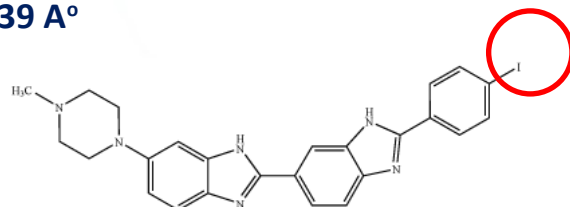
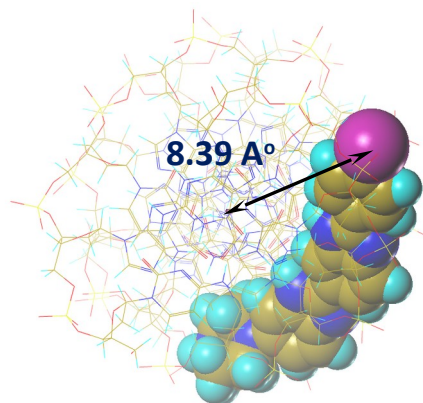
Iodo-Hoechst 33258, $R = 9.12 \text{ Å}$



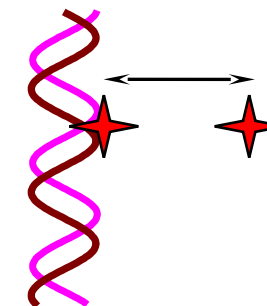
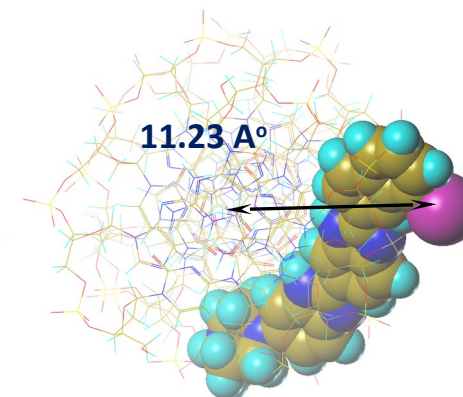
Bi-Benzimidazole-5-Iodo-Naphthyl, $R = 7.41 \text{ Å}$



Para-Iodo-Hoechst, $R = 8.39 \text{ Å}$



Bi-Benzimidazole-8-Iodo-Naphthyl, $R = 11.23 \text{ Å}$

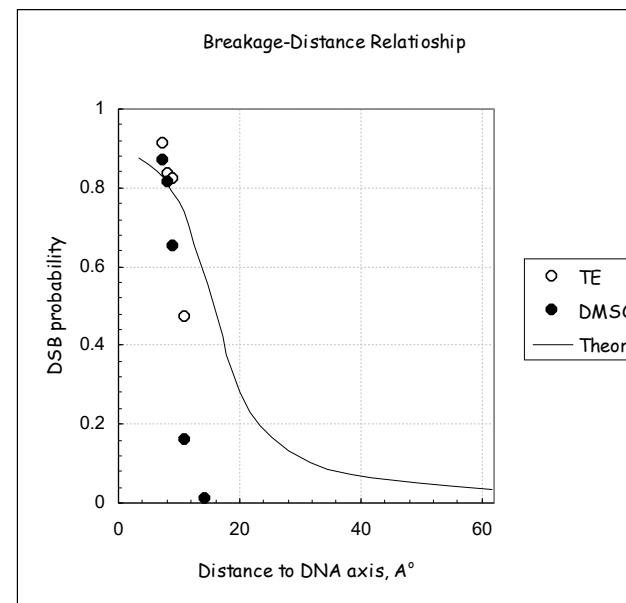
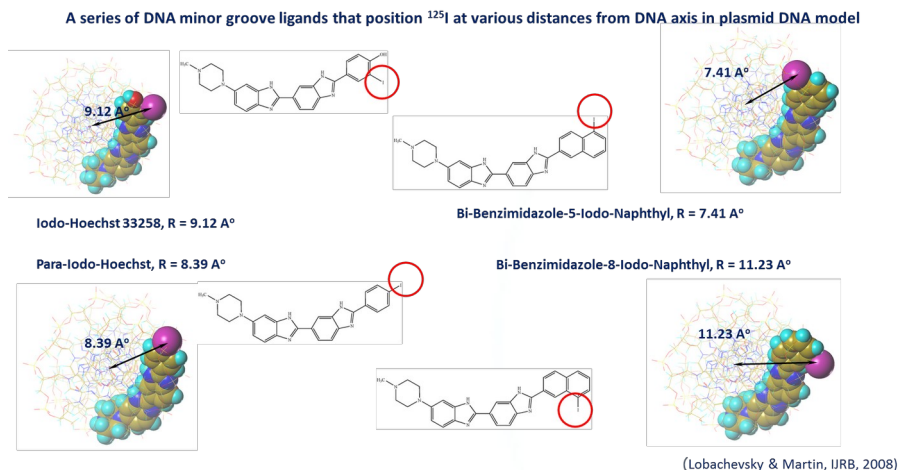


(Lobachevsky & Martin, IJRB, 2008)

Auger decay in DNA ligand – the role of distance



A series of DNA minor groove ligands that position ^{125}I at various distances from DNA axis in plasmid DNA model

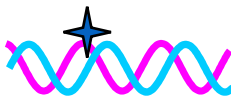


(Lobachevsky & Martin, IJRB, 2008)

Ligand	Distance from ^{125}I to DNA axis	DSB (TE)	DSB (DMSO)
BB5IN	7.41	0.91	0.87
PIH	8.39	0.84	0.81
IH33258	9.12	0.82	0.65
BB8IN	11.22	0.47	0.157

- Decrease of DNA DSB probability with increasing distance to DNA axis
- Distance – critical parameter
- Discrepancy with Monte Carlo result – non-radiation component

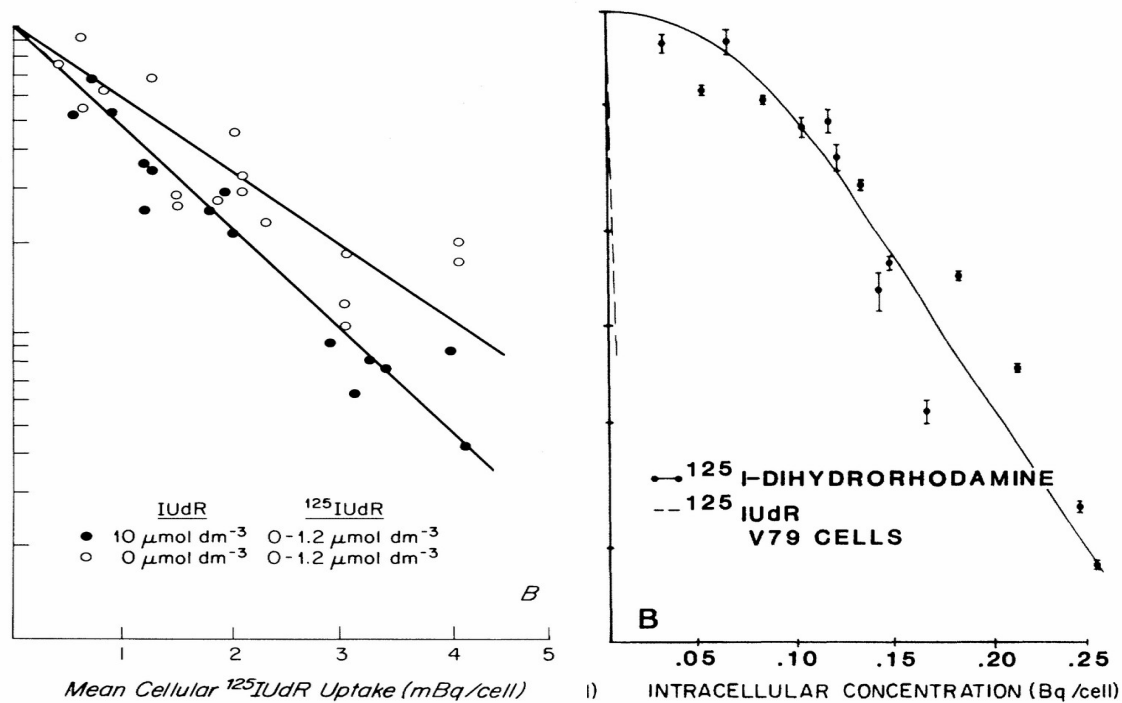
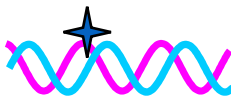
Auger decay in DNA ligand – iodine isotopes ^{123}I , ^{124}I , ^{125}I



	Buffer	^{125}I	^{123}I	^{124}I
Auger electrons (Energy, keV)		24.9 (12.2)	14.9 (7.4)	8.4 (4.74)
DSB (linearisation) /decay	TE	0.82	0.62	0.54
	TE+DMSO	0.65	0.54	0.43
SSB (relaxation) /decay	TE	0.09	0.30	0.40
	TE+DMSO	0.15	0.36	0.41

Decay of “weaker” Auger emitters ^{123}I and ^{124}I is efficient in induction of DNA breaks

Radiation biology of Auger decay – cellular effects



Localisation of Auger emitter relative to DNA is of critical importance

Experiments with mammalian cells (*Kassis & Adelstein, Rad Res, 1987*)

Effect of ^{125}I decay: extracellular, in cytoplasm, in cell nucleus

Decay of DNA incorporated ^{125}I ~ 6 fold more efficient than decay of ^{125}I in cytoplasm (based on absorbed dose)

Carrier of ^{125}I	Localisation	D_{37} mBq/cell	D_{37} Gy
Na^{125}I	Extracellular	-	-
$[^{125}\text{I}]$ -iodo-dihydrorhodamine (^{125}I -DR)	Cytoplasm	109	4.62
$[^{125}\text{I}]$ -iodo-deoxyuridine ($5\text{-}^{125}\text{I}$ UdR)	Nucleus (DNA)	1.30	0.80



1. Auger electron emitters – a special class of radionuclides

(physics of Auger effect and physical properties of Auger emitters)

2. Molecular aspects of Auger decay radiobiology

(DNA damage)

3. Auger emitters potential for radionuclide therapy

(dual targeting strategy)



Properties/Advantages of Auger emitters (iodine-125):

- **Highly localised energy deposition**
- **Efficient induction of DNA damage (complex DSB) by DNA associated decay -> High cytotoxicity of DNA associated radionuclide**
- **Strong dependence on decay-DNA distance**
- **Strong dependence of radiotoxicity on decay localisation (DNA or cytoplasm) presents a potential for selective action on certain cells or cell types without damage to neighbouring cells**
- **Theranostic opportunities – accompanying gamma (SPECT) or positron (PET) emission**

Requirements:

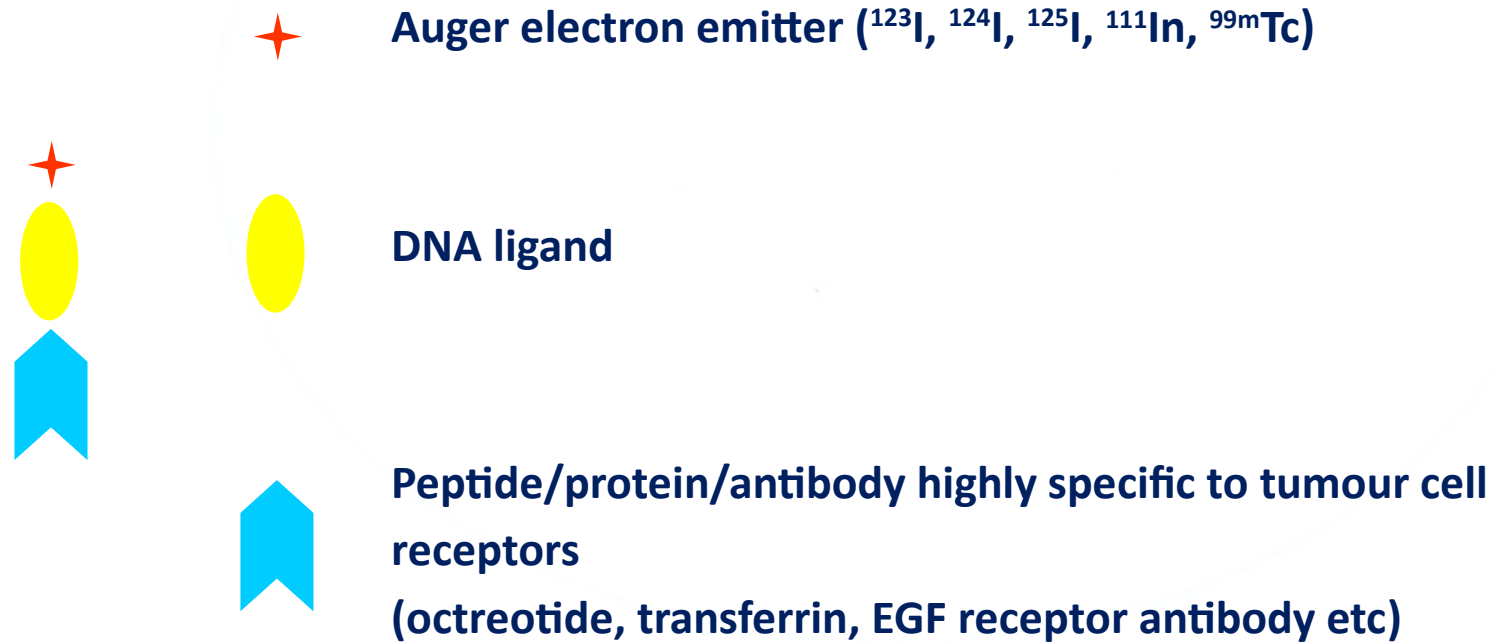
- **- Localisation of Auger decay in close vicinity to cellular DNA**
- **- Selective delivery of Auger emitters to cells of malignant tumours**

Auger emitters potential for radionuclide therapy



Approach – Dual targeting strategy

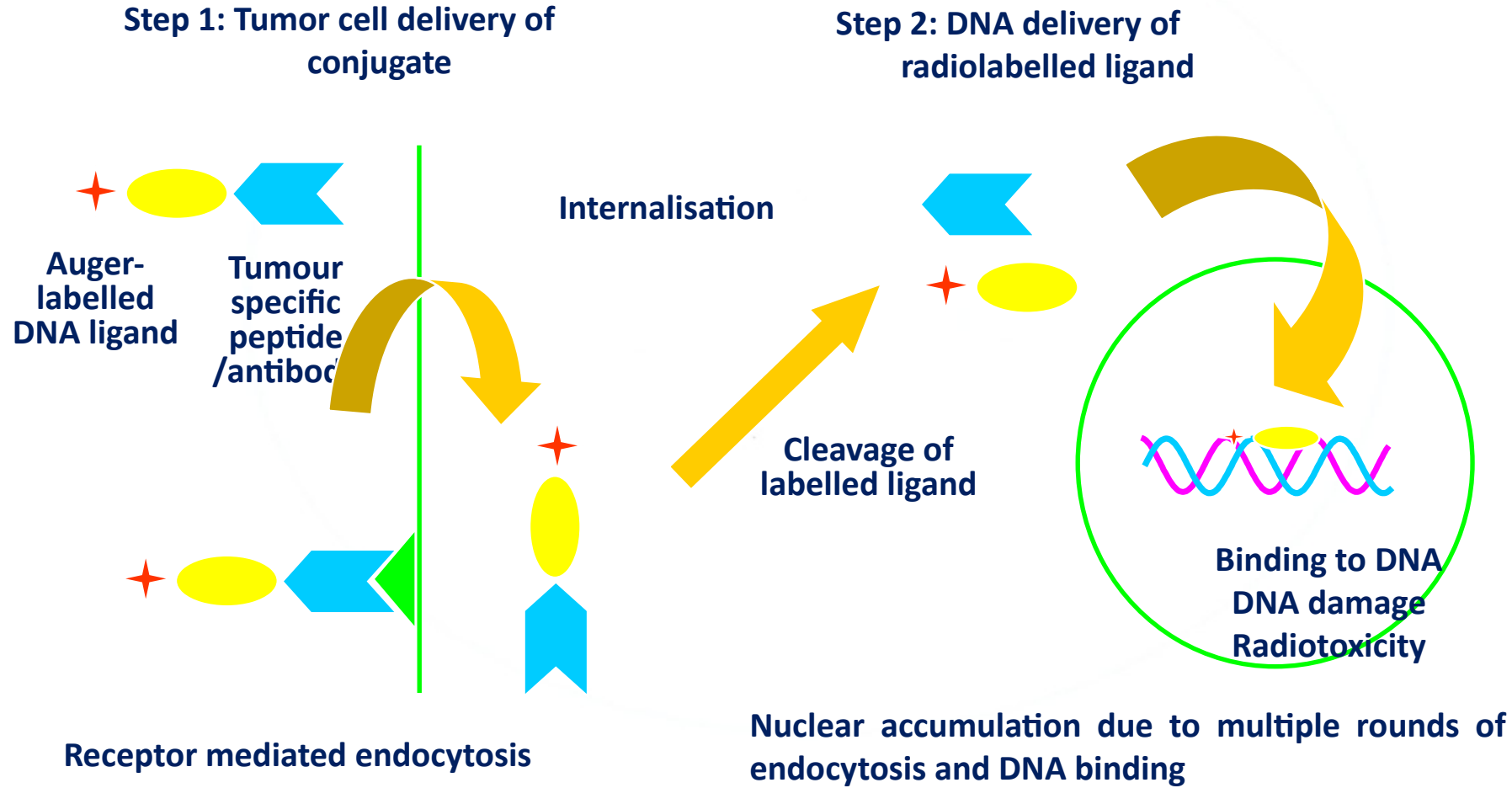
Three-component conjugate – a key element of the strategy
(module transporter)

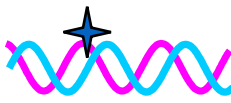


Auger emitters potential for radionuclide therapy



Dual targeting strategy





Dual targeting strategy – Octreotide model

Model – Octreotide (octreotate):

- Octreotide - peptide (8 amino acids) – analogue of somatostatin (growth hormone inhibiting hormone) (14 or 28 amino acids)
- somatostatin receptors (SSTR) are over expressed in many tumours (gastrointestinal, prostate, mammary gland, neuroendocrine)
- radiolabelled Octreotide is used in nuclear medicine

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DOI: 10.3109/09553002.2012.666375

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healthcare

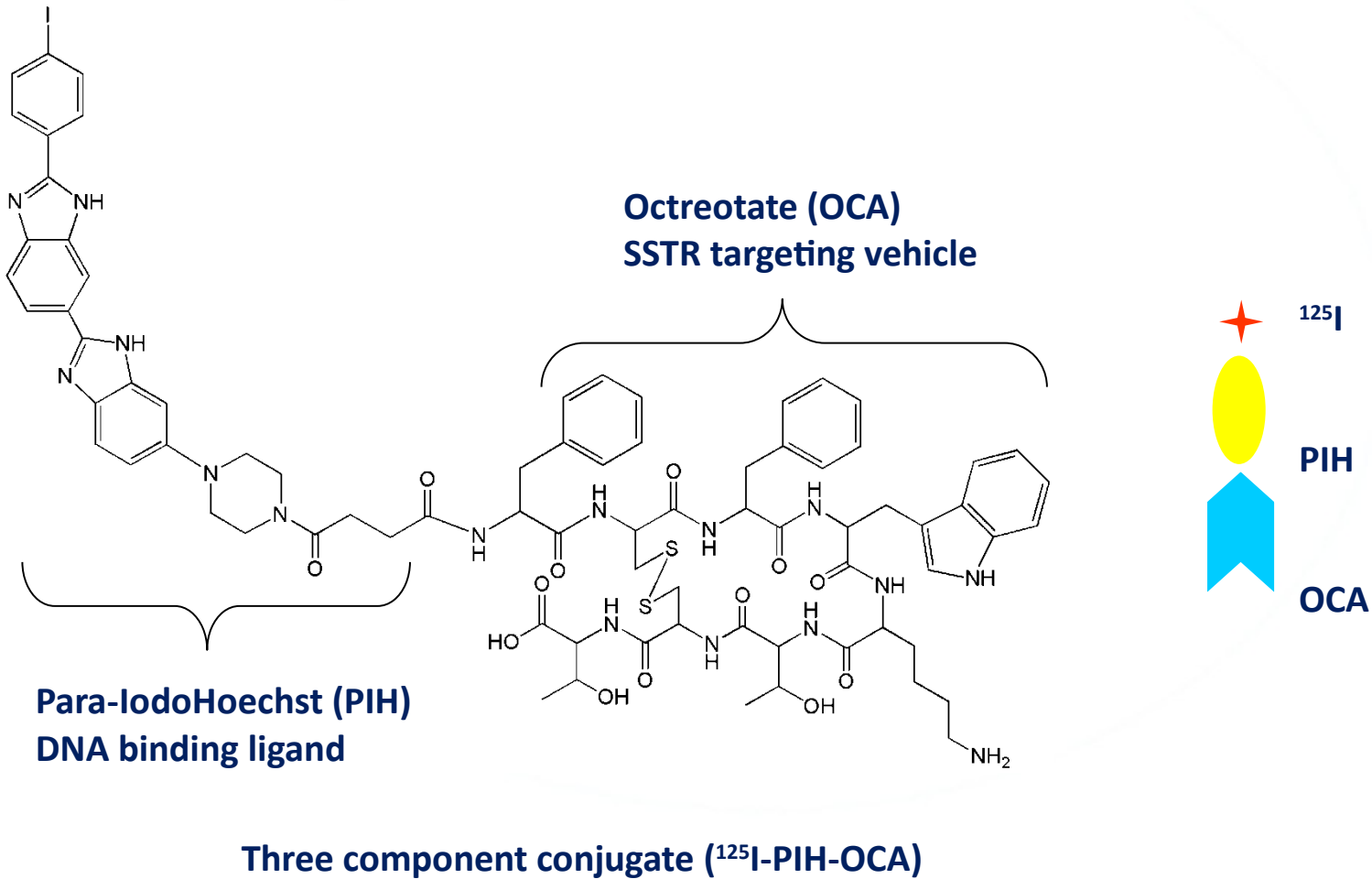
Tumour targeting of Auger emitters using DNA ligands conjugated to octreotate

Pavel Lobachevsky¹, Jai Smith¹, Delphine Denoyer¹, Colin Skene², Jonathan White², Bernard L. Flynn³, Daniel J. Kerr³, Rodney J. Hicks¹ & Roger F. Martin¹

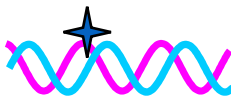
Auger emitters potential for radionuclide therapy



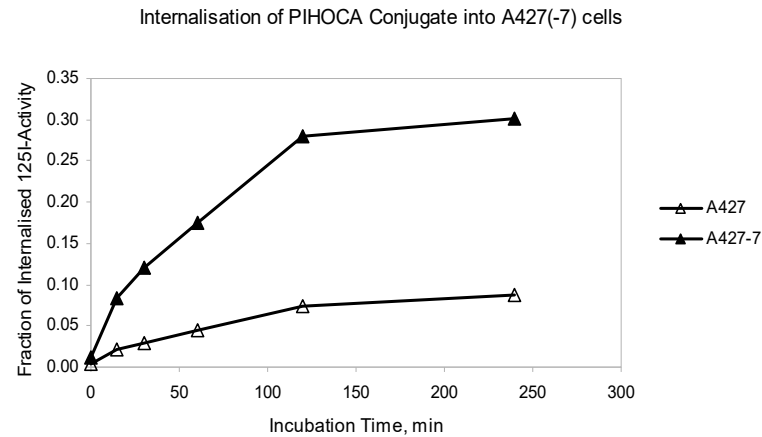
Dual targeting strategy – Octreotide model



Auger emitters potential for radionuclide therapy



Dual targeting strategy – Octreotide model, *in vitro* experiments

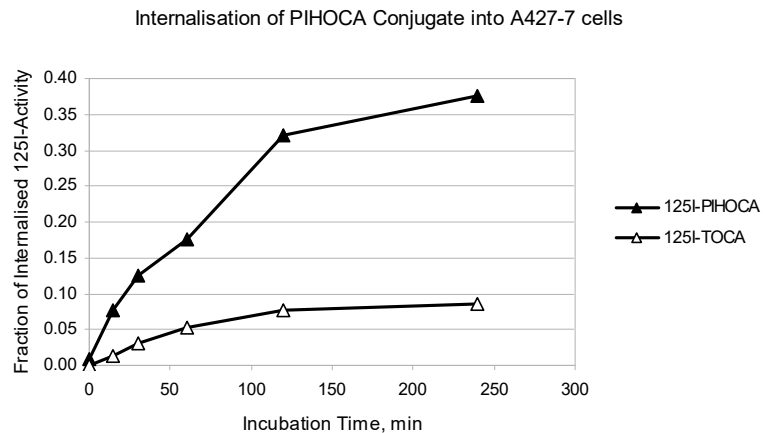
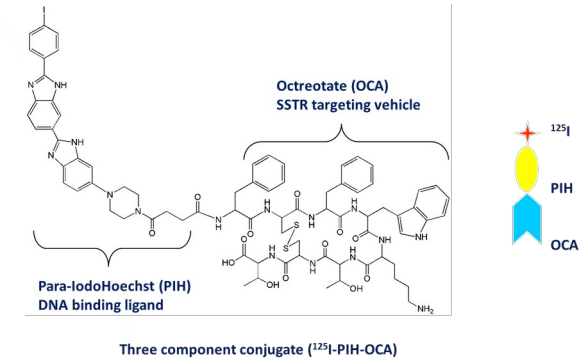


Accumulation of Auger emitter ^{125}I in cells

Open symbols – A427 (parent line)

Black symbols – A427-7 (SSTR2 expressing)

Accumulation is mediated by SSTR2 receptors



Open symbols– directly labelled ^{125}I -TOCA w/o DNA ligand

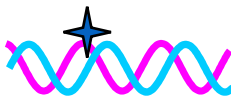
Black symbols – conjugate (^{125}I -PIH-OCA)

Accumulation is mediated by DNA binding of ^{125}I -PIH

Cell culture – A427 (human non-small cell lung carcinoma) and clone

A427-7 stably transfected with somatostatin receptor (SSTR2) gene

Auger emitters potential for radionuclide therapy



Dual targeting strategy – Octreotide model, *in vivo* experiments

^{125}I -PIH-OCA

Biodistribution of Auger emitter in mouse tissues
24 hr after injection

Accumulation of Auger emitter in tumour
following injection of both ^{125}I -PIH-OCA
and ^{125}I -TOCA

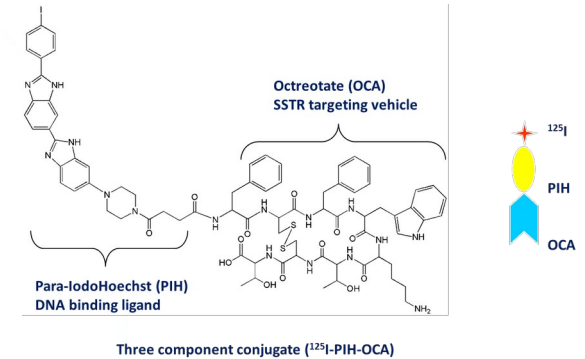
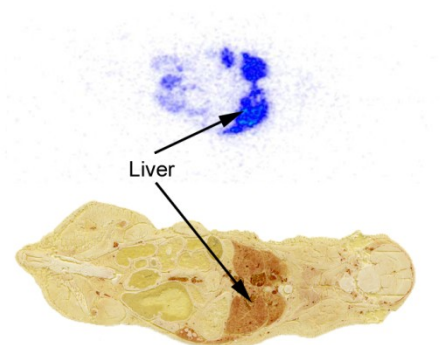
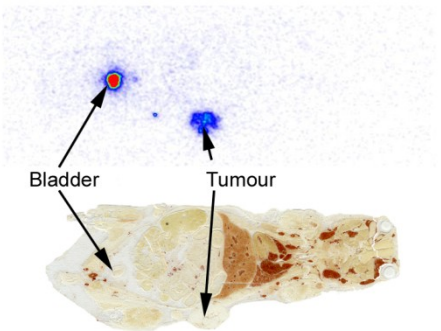
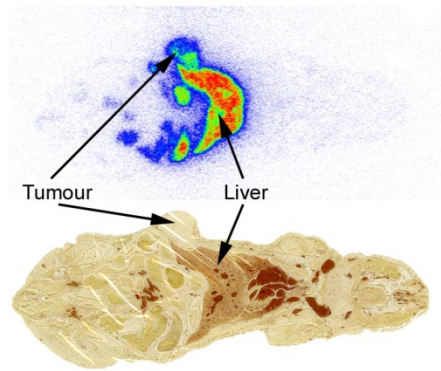
^{125}I -TOCA

Accumulation of Auger emitter in liver following
injection of ^{125}I -PIH-OCA and ^{125}I -PIH

There is targeted delivery of Auger emitter,
however:

^{125}I -PIH

Premature cleavage of DNA ligand and its
accumulation in liver





Dual targeting strategy – Octreotide model

Experimental studies demonstrate the feasibility of targeted delivery of Auger emitters to nucleus and DNA of tumour cells



**Radiobiology of Auger electron emitters
and their potential for radionuclide therapy**

Thank you for your attention!

